

## Bibliometric study of chemical compounds based on medicinal plants anti-Alzheimer



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### **ABSTRACT**

Alzheimer's disease is a chronic neurodegenerative condition that affects mainly memory and other cognitive functions. Interest in finding treatments effective for this disease has increased in recent years, leading to the investigation of the use of medicinal plants and isolated compounds as possible therapeutic treatments. This study aims to identify research on plants and isolated compounds that demonstrate efficacy in the treatment of markers of Alzheimer's pathology over the years.

To achieve this goal, we performed a literature review using methods quantitative, which allows us to identify trends, authors, journals and research areas most relevant to the topic. We collect data and submit them to quantitative analyzes to get an overview of the state of the art in this particular field. The methodology employed included the use of bibliometric software for surveys quantitative surveys and the Rayyan platform for qualitative surveys. The quantitative analysis covered the leadership of the studies, the power analysis used in the articles and the identification of predominant themes. While in the qualitative analysis, we focused on the experimented plants that cause anti-Alzheimer's effects, with a focus on the isolated chemical compounds responsible by pharmacological action. As a result of the quantitative analysis, we found a growing interest in research on plants and Alzheimer's disease. Since the first publications at the beginning of the 2000s to the present day, there has been a steady increase in the number of studies, reflecting a significant progress has been made in understanding the possible benefits of plants in the treatment and in preventing this debilitating disease. Our findings highlight the continuing importance of research in this area, which may provide promising therapeutic alternatives to face the challenges posed by the disease of Alzheimer's. Through this bibliographic research, we hope to contribute to the advancement of scientific knowledge and for the development of more effective therapeutic approaches and accessible for this complex condition.

**Keywords:** Alzheimer's disease, isolated compound, medicinal plants, technology.

## **1 INTRODUCTION**

Alzheimer's disease (AD) is a neurodegenerative pathology discovered in 1906 by the German physician Alois Alzheimer. And as the number of elderly people in the world grows, there is a proportion of new cases of this pathology, due to physiological and immunological factors that

decrease in old age. The main symptom of AD is memory loss, worsening of daily habits, difficulty in understanding, discouragement and mental confusion (Jamshidi-Kia et al., 2018).

For many years human beings have always been in contact with nature and have taken advantage of the environment to obtain tools, supplies and medicines. The ability to use plants as food and to treat diseases was conducted over several failures over time, and progressively man was able to use the flora for his activities. The use of medicinal plants is passed from generation and gradually the understanding of the effectiveness has been increased the reliability of the use of species around the world, due to the potential in the development of drugs favoring public health for the treatment of current and future neurological diseases (Breijeh & Karaman, 2020).

Medicinal plants are a world wealth of importance in food and in the production of new medicines, only in Europe there are about 1300 species of wild plants for use in medicine and in the United States around 118 to 150 of the main existing and prescribed drugs are of natural bases and 80% of developing countries are dependent on herbal drugs. And 25% of developed countries the drugs prescribed are of wild plant origin, this growing consumption by herbal drugs from plants is expanding rapidly around the world in primary health (S.-L. Chen et al., 2016)

According to the Declaration of Alma Ata (1978), primary health care is care based on methodologies, technologies, practices that are reliable and accepted by society, disposed in the community at a cost that everyone can perpetuate from its use and the phases of its development. The declaration of Alma Ata also manifests the primordiality of the incorporation of medicinal plants and phytotherapy in the public health of countries, justifying that 80% of the world population uses plants or preparations from them in primary health (Akram & Nawaz, 2017)

Plant species are made up of healing properties due to the existence of chemical substances called secondary metabolites contained in parts of the plant that have as their purpose self-defense or sustainability in the environment. These metabolites are chemically classified within groups with biological activities flavonoids, alkaloids, anthraquinones, glycosides, tannins, saponins among others and may be present in several species, whether they are land plants, marine plants, lichens and / or fungi capable of assisting in the cure and / or treatment of neurological diseases including Alzheimer's (Panda & Jhanji, 2020)

One of the most important methods employed in the therapy for Alzheimer's disease is the management of normal levels of the enzyme acetylcholine in the synaptic cleft, but these drugs that provide this action have some side and adverse effects to those who consume, so the search for new more efficient drugs is of paramount importance, currently numerous active constituents of plants are explored and experimented in studies as possible drugs in the therapy of neurodegenerative diseases, Due to the availability, lower cost, low toxicological feats of herbal medicines in relation to synthetic drugs make them a better choice. (This approach called ethnopharmacological uses health system and



pathologies and the inclusion of plants with their traditional knowledge, chemical and pharmacological studies in the discovery of new drugs based on the screening of plant extracts or isolated compounds (Tuzimski; Petruczynik, 2022) Pyou; Jhanji, 2020).

The pathology causes increased oxidative damage, neurofibrillary entanglements, decline of the enzyme acetylcholine among other serious problems in the brain that require medications to assist in the health of the person carrying the disease, although there are synthetic and semisynthetic drugs used for AD plants including here those with action on the CNS, especially those with direct or indirect action on the pathology, acting on the mechanism, or in the prevention of disease have importance in health (Panda & Jhanji, 2020).

The review presents studies of medicinal plants and their isolated compounds in the Scopus database effective for Alzheimer's in two methodological stages: qualitative and quantitative, focusing on the production of studies over the years and on the species studied. This study also aims to highlight the most commonly used families and isolated compounds of Anti-Alzheimer's interest.

## **2 ETHNOPHARMACOLOGY AND ALZHEIMER'S DISEASE**

The origins of ethnopharmacology go back to ancient times, when communities depended exclusively on nature to meet their health needs. Over the centuries, these cultures have accumulated a vast knowledge about the medicinal properties of the plants present in their ecosystems. This knowledge was transmitted orally from generation to generation, constituting a true treasure of traditional wisdom and with the advancement of modern science, there was a tendency to devalue this ancestral knowledge. Fortunately, ethnopharmacology has emerged as a discipline to rescue and elevate this knowledge, recognizing its potential in the development of new treatments and therapies (Pirintsos et al., 2022).

One of the main characteristics of ethnopharmacology is its multidisciplinary approach. This area of study combines elements of ethnobotany, ethnology, anthropology, botany, pharmacology and chemistry, among other disciplines. This holistic approach allows for a deeper understanding of traditional practices of medicinal plant use, incorporating cultural, social and scientific aspects (Reyes-Garcia, 2010).

This approach highlights the importance of biodiversity conservation. By investigating medicinal plants valued by local communities, ethnopharmacologists have succeeded in identifying and preserving key species in their natural habitats. This promotes sustainability and conservation of natural resources, ensuring the continued availability of medicinal plants for future generations (Rodrigues et al., 2022) .

One of the main objectives is the discovery of new therapeutics based on traditional knowledge. Many modern medicines have their origins in compounds found in plants used in



ethnopharmacological practices. Research in this area can lead to the identification of new bioactive compounds, providing knowledge for the development of more effective treatments with fewer side effects (Turpin et al., 2022).

This approach has shown promise in the context of Alzheimer's disease, as many cultures possess ancestral knowledge about using plants to promote cognitive health and improve brain function. Understanding the ethnopharmacological practices related to Alzheimer's disease is critical to identifying plants with potential neuroprotective or cognitive-enhancing properties. Several cultures around the world have used medicinal plants as part of their traditional treatments for impaired memory and brain function problems (Gregory et al., 2021).

Through scientific research, it is possible to investigate the chemical properties of plants and identify bioactive compounds that can act in the brain, protecting nerve cells and accepting the neuroinflammatory processes associated with Alzheimer's disease. In addition, ethnopharmacology values the traditional knowledge and ancestral wisdom of indigenous communities. This information is valuable for the discovery of new therapies and may open avenues for the development of drugs for neurological diseases that are more effective and have fewer side effects (Grodzicki & Dziendzikowska, 2020).

A notable example is the Ginkgo biloba plant, widely used in traditional Chinese medicine and other Asian cultures. Ethnopharmacological and scientific studies have shown that Ginkgo biloba extract has antioxidant and anti-inflammatory properties, which can improve cognitive function and slow the progression of Alzheimer's disease (Tewari et al., 2018a).

Another plant that arouses interest in ethnopharmacology is turmeric (*Curcuma longa*), known for its anti-inflammatory and antioxidant action. Turmeric's main active compound, curcumin, has been the subject of research for the treatment and prevention of Alzheimer's disease due to its neuroprotective properties and its ability to inhibit the formation of beta-amyloid protein plaques in the brain, a hallmark of the disease (Mishra & Palanivelu, 2008; Sharifi-Rad et al., 2020).

Ethnopharmacology also contributes to the discovery of new therapeutic targets. Through the investigation of ethnopharmacological practices, it is possible to identify plants and substances that operate in different molecular pathways related to Alzheimer's disease, such as the regulation of beta-amyloid metabolism, the fight against oxidative stress and the modulation of brain inflammation (Tewari et al., 2018b).

Although ethnopharmacology has guarantees for Alzheimer's disease research, it is important to note that studies in this area are still ongoing. Scientific validation of these medicinal plants and natural substances as effective treatments requires rigorous investigation and further clinical studies (Akram & Nawaz, 2017)



In short, ethnopharmacology plays a key role in Alzheimer's disease research, exploring the traditional knowledge of different cultures and identifying medicinal plants with therapeutic potential. This multidisciplinary approach offers new perspectives and could lead to the development of innovative and more effective treatments for this neurodegenerative condition that affects millions of people around the world (Tyler & Tyler, 2023).

### 3 METHODOLOGIES

In order to analyze the production of scientific studies on plants and isolated compounds for Alzheimer's disease, we opted for bibliometric research by analysis of the Software-RStudio through the tool Bibliometrix and Scopus. (Aria & Cuccurullo, 2017).

Bibliometrics is an area that addresses quantitative research, analysis of several studies detailed as year of publication, countries, journals, most cited authors, funders among others, this series of evidence shows the importance of scientific production in various areas such as health, humanities, education, technologies among others. Through statistical data, bibliometrics makes it possible to evaluate and monitor the evolution of the field of study through the classification of citations and cooperation through keywords and themes. As a result, one can know about the area (GUIMARAES; MOREIRA; BEZERRA, 2021).

For the synthesis of the first data, the date of collection on February 14, 2023 of the documents was chosen; choice of key terms by applying the Boolean operators TITLE-ABS-KEY (isolated AND compound AND alzheimer's AND plants); collection of bibliometric data in the Scopus database in which there was no exclusion of language or temporal design, having 530 records, then imported the documents in "BibTeX" format. Then, installed the RStudio Software - Applying the codes ("BIBLIOMETRIX", "LIBRARY (bibliometrix)", "BIBLIOSHINY ()"), in order to access the bibliometrix package and insert the documents in "BibTeX" format. To evaluate the quantitative analysis of research carried out over time, analysis of terminologies among authors and thematic evaluation among authors.

In the second step of data extraction applying the same key terms and filter in the Scopus database, articles from reviews, book chapters, short researches, conferences, books, editorials, letters and observations were excluded. There are 399 documents left for analysis that were inserted in the tool (Rayyan) - AI Powered Tool for Systematic Literature Reviews that enables the qualitative development of studies.

Of 399 documents 212 were included. And for the insertion of these 212 researches was read the abstracts and only the isolated active compounds with positive results in which the terms mentioned by the authors such as "first report", "potent inhibition", "more potent", "more active", "significantly", "strong inhibitor", "potently", "higher activity", "more effective activity", "more promising".



The 187 excluded articles had isolated substances with moderate and weak effect, studies that included seaweed, activity in neuropathological diseases that did not exclusively encompass Alzheimer's disease, inhibition by extract, did not mention the plant, studies of fungi, bacteria, review articles, no anti-Alzheimer's action or anti-Alzheimer's activity.

## 4 OUTCOME AND DISCUSSION

### 4.1 PRODUCTIVITY ANALYSIS

The first work produced regarding the keywords inserted in the Scopus database is authored by researchers Jin-Hui Kim, Sang-In Kim and Kyung-Sik Song, published in 2001. Under the title *Prolyl Endopeptidase inhibitors from green tea*, research conducted by methanolic extract of green tea leaves succeeded in isolating three compounds gallate from (-)-epigallocatechin, gallate from (-)-epicatechin and gallate from (+)-gallocatechin. From 2003 to 2006 had a continuous growth, the other years until the realization of this research it is possible to verify several oscillations. Relating to the measures taken worldwide of social isolation that have driven many researchers away from their face-to-face studies due to Covid-19, the year 2020 presented a satisfactory rate of documents with 62 publications compared to 2022 which obtained 69.

The most productive author according to Scopus' metric analysis is Choi, J. S. has 17 announced papers addressing studies of plants, Alzheimer's disease, cholinergic inhibition and neuroprotection. The first research conducted by the author entitled *Cholinesterase and BACE1 inhibitory diterpenoids from Aralia cordata* in the year 2009. The researcher has as first author 6 articles published in the *Archives of Pharmacal Research*; the others divided among other sources as co-author.

In analysis of publication of the documents by funding institution highlighted by Scopus China has (127 articles) and in second place South Korea (86 articles) until the date of this research, countries such as India, Japan, Spain, Saudi Arabia, Egypt, Pakistan, United States and Brazil do not appear as major producers by funding foundation, however by country / territory all other countries mentioned above, are the 10 largest producers of articles published on the plant over the years, where each one presents more than 20 indexed documents. Looking at the time frame from 2011 to 2017, the small difference between the two largest producers: China with 48 articles and South Korea presenting 40 plant research articles.

This number of studies involving plants and Alzheimer's disease may be related to the number of elderly people living in this country. China is the second most populous country in the world second only to India and according to China's seventh national census in 2020, there are 264,018,766 and 190,635,280 elderly people aged 60 and 65, if you buy with census conducted in 2010 the country's elderly population has aged rapidly and the number of incidences, mortality and problems related to

aging gradually increases. The existence and incidence of the increase in Alzheimer's disease presents several problems in public health and in the urban and rural society of China, this recent census points out that there are 15.07 million elderly people with dementia and of this totality 9.83 million are with Alzheimer's. A study also showed that China spent US\$ 167.74 billion on treatment for people with Alzheimer's in the year 2015 and points out that by the year 2050 the costs of this pathology may reach 1.8 trillion (R. Ren et al., 2022)

#### 4.2 ANALYSIS OF VOCABULARIES

In the evaluation of the keywords used by all the researchers, inserted in the Bibliometrix tool, it was found 2758 sets of words where the term "Alzheimer's disease" was the most assiduous, with 404 appearances, in second place, word "plant extract", which repeats twice, as shown in Table 1. This denotes the importance of plant studies and a network of occurrence of this term during the passage of time as shown in Figure 3.

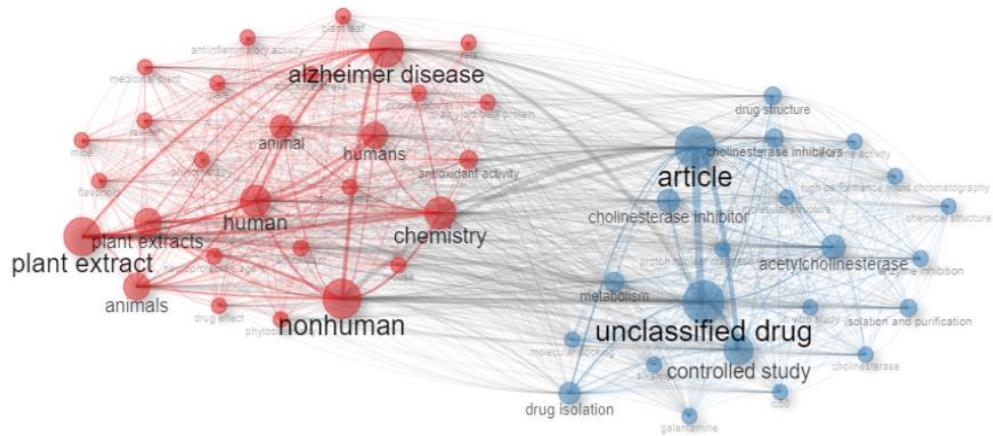
Table 1. Frequency, keywords and consistencies

Words	Occurrences
Alzheimer's disease	404
Plant Extract	325
Unclassified drugs	308
Articles	304
Non-Human	287
Acetylcolinesterase	266
Human	244
Química	224
Control Study	202
Plant extract	194

Source: Adapted Bibliometrix (2023)

Figure 3 represents a set of ideas between authors and co-authors of related studies. The lines show a link between the keywords mentioned by the authors. On the red side, the featured keywords are "Alzheimer's disease," "plant extract," "human," "animal," "chemical," and "non-human," indicating a research collaboration over time among researchers in these areas. On the blue side, there are two featured terms, "articles" and "unclassified drug," which appear to be related to observational research studies and data collection. This figure can represent the interconnection of ideas and collaboration between researchers in different areas of study, as well as the collection of data through observational studies.

Figure 3. Co-occurrence Network

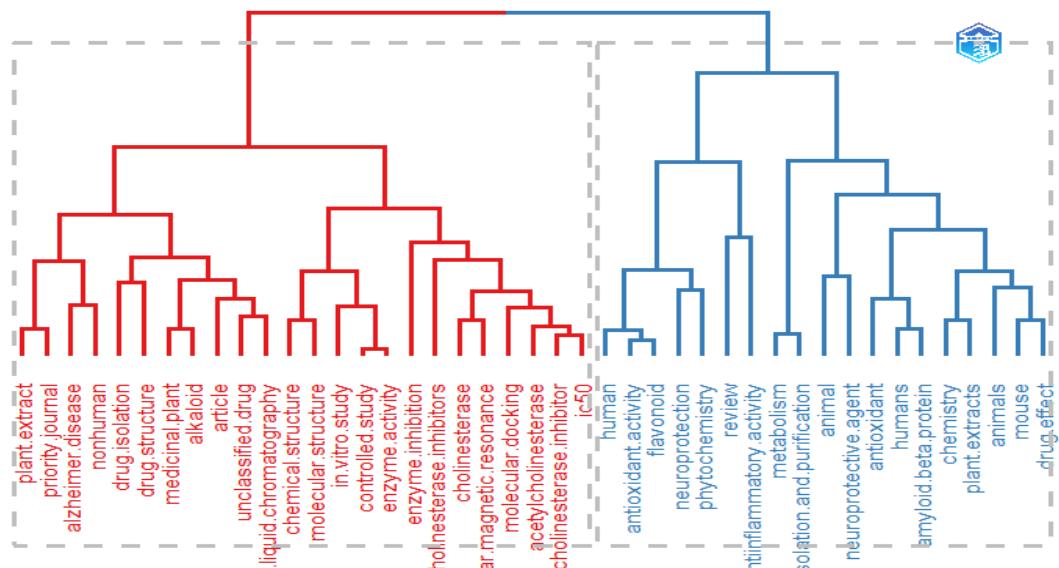


Source: Bibliometrix

#### 4.3 EVALUATION BY THEME

In the analysis of the themes more in accordance with (Figure 4), it is possible to observe that the most recurrent themes in the research of plants and alzheimer are formed by 2 clusters in blue and red color.

Figure 4. Dendrogram



Source: Bibliometrix (2023)

Figure 4 shows two clusters, one in blue and the other in red, which represent the most recurrent themes in plant and Alzheimer's research. Through the analysis of the dendrogram, it is possible to observe the order and connections between these most frequent themes.



In total, 24 central themes with a high degree of development are identified, which are related to phytochemical studies of plants, isolation of chemical compounds and neurological inhibitory biological activities, both in humans and in non-humans.

The themes highlighted in blue are related to 19 central themes and represent studies in humans, isolation of compounds, antioxidant potential and control studies. In the dendrogram, it is possible to observe that the use of plants is related to all areas, from the tolerance and isolation of active substances to the performance of tests to verify the effects of drugs.

This analysis suggests a strong connection between plant phytochemical studies and Alzheimer's disease research, with a wide range of related topics and a comprehensive approach from administration to evaluation of the therapeutic effects of isolated compounds.

#### 4.4 MEDICINAL PLANTS INHIBITING THE ENZYMES ACETYLCHOLINESTASIS (ACHE) AND BUTYRYLCHOLINESTERASE (BUCHE)

One of the most widely used methods for detecting enzymatic inhibitory activity is the Ellman technique. This test is performed by means of photometry that has the ability to cause action of the enzyme through tissue extracts, homogenized fractions, isolated compounds among others. In this test the enzymatic activity is observed by the growth of the yellow color that is formed due to thiocholine, this occurs at the moment that thiocholine comes into contact with the reagent of 5,5'-dithiobis-(2-nitrobenzoic acid) or DTNB) the production of anions of yellowish color. This same enzymatic method written by Ellman et al, is also used with alteration by. That includes the thin-layer chromatography (CCD) technique. The studies by Thin Layer Chromatography are performed to trace plant extracts prepared in an amount of 10mg/mL added in a certain chromatographic plate in a thin layer of silica gel (Ellman et al., 1961) (Rhee et al., 2003) (Castro and Silva et al., 2020)

In addition to this enzymatic study several researchers use *in vivo method by the tissue* of the frontal cortex, in the cerebellum even in the area of memory, this method has the objective of observing improvements in memorization, inhibitory protection induced by oxidative stress among others (Farias et al., 2022)

Existing treatments for Alzheimer's disease use drugs to increase acetylcholine (ACh) levels, causing inhibition of acetylcholinesterase and butyrylcholinesterase that are in charge of hydrolyzing ACh into acetate and choline, act only in the progressive relief of Alzheimer's (Masondo et al., 2019)

In the brains of people diagnosed with Alzheimer's, the enzyme acetylcholinesterase is found in large proportion more than butyrylcholinesterase, this increase causes the rapid hydrolysis of acetylcholine prevented the action of the neurotransmitter (Marucci et al., 2021)

The importance of compounds presents in plants called secondary metabolites in addition to helping in the survival and protection of plants, has become fundamental for humans in the

development of medicines. And these chemicals extracted from plants that have an affinity to act on the central nervous system (CNS) are of interest for the development of new pharmaceutical drugs as shown in (table 2) studies that obtained inhibitory activity of enzymes for Alzheimer's disease carried out over the years.

Tables 2. Medicinal plants with anticholinesterase and anti-butryrylcholinesteratic activity

Plants and Families	Anti-Alzheimer's Compounds	Author
<i>Piper nigrum L.</i> (Piperaceae)	Piperine (derivative)	(Jaipea et al., 2023)
<i>Cassia timoriensis DC.</i> (Fabaceae) <i>Cassia grandis Lf</i> (Fabaceae)	luteolin, cinnamic acid, 4-hydroxycinnamic acid	(Alhawarri et al., 2023)
<i>Grewia tiliaefolia Vahl</i> (Malvaceae)	propyl 3-hydroxy-10,13-dimethyl-6,7,8,9,10,11,12,13,14,15,16,17-dodecahydro-3H-cyclopenta [a]phenanthreno-17-carboxylato	(Rajput et al., 2023)
<i>Fragaria ananassa Duchesne</i> (Rosaceae)	4,22-cholestadien-3-one, stigmast-4-en-3-one	(Mahnashi & Alshehri, 2022)
<i>Andrographis paniculata</i> (Burm.f.) Wall. (Acanthaceae)	desoxy-Andrographolide (Derivative)	(Jatav et al., 2022)
<i>Fritillaria taipaiensis P.Y.Li</i> (Liliaceae)	taipainin (Derivative)	(A.-W. Wang et al., 2022)
<i>Talguenea quinquenviria</i> (Gillies & Hook.) I.M. Johnst., (Rhamnaceae)	Zizibberenalic acid, zizibberanalic acid, ceanotic acid, ceanotenic acid, ceanotannolic acid, 3-oxo-oceanic acid, 3-O-acetyl-oceanic acid	(Muñoz-Nuñez et al., 2022)
<i>Notholirion thomsonianum</i> (Royle) Stapf (Liliaceae)	2-(3,4-dimethoxifenyl) -3,7-dihidroxi-4H-cromen-4-one	(Mahnashi, Alshahrani, et al., 2022)
<i>Plantago subulata L.</i> (Plantaginaceae)	acteoside, isoacteoside, echinacoside, and arenarioside	(Özaslan et al., 2022)
<i>Polygonum aviculare L.</i> (Polygonaceae)	hydroxy succinimide	(Mahnashi, Alyami, et al., 2022)
<i>Pongamia pinnata</i> (L.) Pierre (Fabaceae)	pongapine, ovalichromene B, gamatin and pongaglabrone	(Nguyen et al., 2022)
<i>Vincetoxicum funebre</i> Boiss. & KotschyName (Apocynaceae)	daucosterol	(Abbas-Mohammadi et al., 2022a)
<i>Conocarpus lancifolius Engl</i> (Combretaceae)	lancifolamide	(Saadullah et al., 2022)
<i>Berberis parkeriana CKSevy.</i> (Berberidaceae)	3-O-(p-Bromobenzoyl)jatrorrhizine	(R. Ali et al., 2022)
<i>Caragana balchaschensis</i> (Kasn. ex Kom.) Pojark. (Fabaceae)	quercetin (derivatives)	(Zhumanova et al., 2021)
<i>Castanopsis cuspidata</i> var. <i>sieboldii</i> (Makino) Nakai (Fabaceae)	4'-O-( $\alpha$ -L-rhamnopyranosyl) -3,3',4-tri-O-methylillagic acid and 3,3',4-tri-O-methylillagic acid	(Oh et al., 2021)



<i>Spiranthes sinensis</i> (Pers.) Ames (Orchidaceae)	quercetin, kaempferol, 3-(4-tolyloxy)-propanolic acid, ethyl ferulate	(Zou et al., 2021)
<i>Mitragyna speciosa</i> Korth. (Rubiaceae)	mitragynine	(Innok et al., 2021)
<i>Vanda roxburghii</i> R.Br. (Orchidaceae)	Gigantol, syringaldehyde	(Ahammed et al., 2021)
<i>Grewia optiva</i> J.R. Drumm. ex Burret (Malvaceae)	2,2'-(1,4-phenylene) bis (3-methylbutanoic acid)	(Ul Bari et al., 2021)
<i>Lawsonia inermis</i> var. <i>spinosa</i> (L.) Pers. (Lythraceae)	3-O-β-acetyloleanolic acid, oleanolic acid	(Balaei-Kahnamoei et al., 2021)
<i>Astragalus membranaceus</i> fish. ex Bunge (Fabaceae)	calycosin-7-O-β-d-glucoside, pratensein-7-O-β-d-glucoside, formononetin-7-O-β-d-glucoside, calycosin, genistein and formononetin	(S. Li et al., 2021)
<i>Cremastra appendiculata</i> (D. Don) Makino (Orchidaceae)	cremaphenanthrene	(L. Liu et al., 2021)
<i>Amaryllis belladonna</i> E. Mey. ex Steud. (Amaryllidaceae)	Acetylarnosine	(Sibanyoni et al., 2020)
<i>Rauvolfia vomitoria</i> Wennberg (Apocynaceae)	vobasenal	(Zhan et al., 2020)
<i>Acacia auriculiformis</i> A. CUNN Ex. Benth (Fabaceae)	α-spinasterol	(Lawal et al., 2020)
<i>Carissa carandas</i> L. (Apocynaceae)	1-Heneicosanol; N-nonadecanol-1; cholesta-4,6-dien-3-ol, (3beta); di-n-octyl phthalate; 7,9-di-tert-butyl-1-oxaspiro (4,5) deca-6,9-diene-2,8-dione; 6-undecyl-5,6-di-hydro-2H-pyran-2-one e phenol, 2,4-di-t-butyl-6-nitro	(Kareti & Pharm, 2020)
<i>Narcissus tazetta</i> var. <i>orientalis</i> (L.) hört. (Amaryllidaceae)	11-Hydroxygalanthin and narcissidin	(Karakoyun et al., 2020)
<i>Piper longum</i> L. (Piperaceae)	piperine	(Khatami et al., 2020)
<i>Hedyotis spread</i> Spreng. (Rubiaceae)	6-O-E-pcoumaroyl scandoside methyl ester, quercetin-3-O-[2"-O-(6"-O-O-E-feruloyl) -β-d-glucopyranosyl] -β-d-glucopyranoside, E-6-O-feruloyl scandoside methyl ester, 6-O-methyldeacetylasperulosidic acid methyl ester, asperulosidic acid, deacetylasperulosidic acid methyl ester, scandoside methyl ester, 6-O-methylscandoside methyl ester.	(J. H. Park & Whang, 2020)
<i>Hippeastrum reticulatum</i> Herb (Amaryllidaceae)	N-chloromethylnarcissidinium, narciprimine, N-methyltyramine, 3β,11α-dihydroxy-1,2-dehydrocrinane	(Hoang et al., 2020)
<i>Tinospora cordifolia</i> (Willd.) Miers ex Hook.f. & Thomson (Menispermaceae)	Oxoglaucone, liriodenine, N-formylanonein	(Onoja et al., 2020)
<i>Erythrina caffra</i> White (Fabaceae)	Erythralin, erythrinin, cristinin A	(Nassief et al., 2020)
<i>Zanthoxylum rigidum</i> Humb. & Bonpl. ex Willd. (Rutaceae)	avicin	(Gonzalez et al., 2020)
<i>Leucophyllum ambiguum</i> Bonpl. (Scrophulariaceae)	furofuranone (derivatives)	(Rios et al., 2020)

<i>Anarrhinum pubescens</i> Loudon ( <i>Plantaginaceae</i> )	iridoids (derivatives)	(Mahran et al., 2020)
<i>Narcissus pseudonarcissus</i> subsp. <i>bicolor</i> (L.) Baker ( <i>Amaryllidaceae</i> )	carltonin	(Al Mamun et al., 2020)
<i>Geophila repens</i> (L.) I.M. Johnst ( <i>Rubiaceae</i> )	pentylcurcumene	(Dash et al., 2019)
<i>Andrographis paniculata</i> (Burm.f.) Wall. ( <i>Acanthaceae</i> )	3,4-di-di-o-caffeoylequinic acid, apigenin and 7-o-methylwogonin	(Benche et al., 2019)
<i>Anthocleista vogelii</i> Planch. ( <i>Gentianaceae</i> )	swertisine	(Ajayi et al., 2019)
<i>Dioscorea communis</i> (L.) Caddick & Wilkin ( <i>Dioscoreaceae</i> )	2,4,8-Trimethoxy-3,7-phenanthenediol	( Boudjada et al., 2019)
<i>Zanthoxylum bungeanum</i> Maxim. ( <i>Rutaceae</i> )	3,4-Dihydroxyphenylethanol	(C.-H. Li et al., 2019)
<i>Solenostemma argel</i> (Delile) Hayne ( <i>Apocynaceae</i> )	kaempferol	( Demmak et al., 2019)
<i>Atalantia monophylla</i> (L.) DC ( <i>Rutaceae</i> )	lupalbigenin	(Posri et al., 2019)
<i>Elsholtzia ciliata</i> (Thunb.) Hyl. ( <i>Lamiaceae</i> )	acacetin (derivatives)	(Nugroho et al., 2019)
<i>Berberis vulgaris</i> L. ( <i>Berberidaceae</i> )	aromoline	(Hostalkova et al., 2019)
<i>Artocarpus lakoocha</i> Roxb. ( <i>Moracea</i> )	2-Arylbenzofurans (derivatives)	(Namdaung et al., 2018)
<i>Artobotrys spinosus</i> Craib ( <i>Annonaceae</i> )	O-Methylmoschatoline, artacinatine C	(Sichaem et al., 2018)
<i>Narcissus poeticus</i> L. ( <i>Amaryllidaceae</i> )	narcipavine	(Šafratová et al., 2018)
<i>Cassia obtusifolia</i> L. ( <i>Fabaceae</i> )	rubrofusarin 6-O- $\beta$ -D-gentiobioside, nor-rubrofusarin 6-O- $\beta$ -D-glycoside	(Shrestha et al., 2018)
<i>Tephrosia purpurea</i> (L.) Pers. ( <i>Fabaceae</i> )	trans-tefrostacin	(Pitchai et al., 2018)
<i>Delphinium denudatum</i> Wall. ( <i>Ranunculaceae</i> )	Norditerpenoid	(H. Ahmad et al., 2018)
<i>Hieronymiella marginata</i> (Pax) Hunz. ( <i>Amaryllidaceae</i> )	sanguinine	(Ortiz et al., 2018)
<i>Stachys japonica</i> Miq. ( <i>Lamiaceae</i> )	4'-O-methylisoscutellarein 7-O-(6"-O-acetyl)- $\beta$ -D-allopyranosyl (1 $\rightarrow$ 2)- $\beta$ -D-glucopyranoside	(Nugroho et al., 2018)
<i>Polygonum limbatum</i> Meisn. ( <i>Polygonaceae</i> ) <i>Dorstenia barteri</i> Bureau ( <i>Moracea</i> )	(-) pinostrobin, 2',4'-dihydroxy-3',6'-dimethoxychalcone, 6-8-diprenyleriodictyol, isobavachalcone, 4-hydroxylonchocarpine e 6-prenylapigenin	(Dzoyem et al., 2017)
<i>Leonurus japonicus</i> Miq. ( <i>Lamiaceae</i> )	caffeic acid, quercetin, p-coumaric acid, kaempferol and hydroxytyrosol.	(Nugroho et al., 2017)

<i>Rosmarinus officinalis (l.)</i> (Lamiaceae)	Nepitrin	(Karim et al., 2017)
<i>Cornus officinalis Siebold &amp; Zucc.</i> (Cornaceae)	telimagrandin II, 1,2,3,6-tetra-O-galloyl-β-d-glucose	(Bhakta et al., 2017)
<i>Morinda officinalis F.C. How</i> (Rubiaceae)	anthraquinones, coumarin (derivatives)	(Y. K. Lee et al., 2017)
<i>Pueraria lobata</i> (Fabaceae)	Lupeol	(Koirala et al., 2017)
<i>Kaempferia parviflora Wall. ex Baker</i> (Zingiberaceae)	3,5,7,3',4'-pentamethoxyflavone	(Seo et al., 2017)
<i>Argemone platyceras Link &amp; Otto</i> (Papaveraceae)	(-)munitagin	(Grid et al., 2017)
<i>Rhodiola crenulata</i> (Hook.f. & Thomson) <i>H. Ohba</i> (Crassulaceae)	(-)Epicatechin gallate ((-)ECG), rhodionin, herbacetin, rhodiosin	(F.-J. Li et al., 2017)
<i>Carthamus tinctorius L.</i> (Asteraceae)	carthatins	(Peng et al., 2017)
<i>Juncus acutus L.</i> (Juncaceae)	Juncunol	(M. J. Rodrigues et al., 2017)
<i>Mutellina purpurea</i> (Poir.) Reduron, <i>Charpin &amp; Pimenov</i> (Apiaceae)	Pteryxin	(Orhan, Senol, et al., 2017)
<i>Fritillaria walujewii Regel</i> (Liliaceae)	tortifolin, Walujewine C, Sinpeanine A, Walujewine E, Walujewine A,	(Y.-M. Liu et al., 2017)
<i>Crataegus oxyacantha</i> Walter (Rosaceae)	β-Sitosterol-3-O-β-D-Glucopyranoside	(M. Ali et al., 2017)
<i>Salsola grandis</i> Freitag, Vural & Adigüzel (Amaranthaceae)	N-acetyltryptophan	(Orhan, Kucukboyaci, et al., 2017)
<i>Valeriana officinalis</i> var. <i>latifolia</i> Briq. (Caprifoliaceae)	volvalerenic acid K	(H.-W. Chen et al., 2016)
<i>Mesua congestiflora</i> (Clusiaceae)	α-mangostin, congestiflorone acetate	(Teh et al., 2016)
<i>Pycnanthus angolensis</i> (Welw.) Campaigne (Myristicaceae)	pongaflavone, (2R,3R)-3-hydroxy-5-methoxy-2",2"-dimethylpyrano [7,8:5",6"]-flavanone	(Elufioye et al., 2016)
<i>Xylia xylocarpa</i> (Roxb.) W. Theob. (Fabaceae)	(3b)-hopan-3-ol-28,22-olide , lupeol, betulin, 28-norlup-20(29)-ene-3β-hydroxy-17β-hydroperoxide, betulinic acid, oleanolic acid, 3β-formyloxy-18α-oleanane-28,19β-lactone, 2,6-dimethoxy-p-benzoquinone	(Lam et al., 2016)
<i>Cryptocarya infectoria</i> Miq. (Lauraceae)	2-Methoxy-atherosperminine	(Wan Othman et al., 2016)
<i>Hoppea fastigiata</i> (Griseb.) C.B. Clarke in Hook.f. (Gentianaceae)	1,5,7-tri-hydroxy-3-methoxyxyanthione, 1,5-di-hydroxy-3,7-dimethoxyxyanthone, 1,3,5-tri-hydroxy-8-methoxyxyanthione	(Moon et al., 2015)
<i>Ampelopsis brevipedunculata</i>	santin, 7-O-α-rhamnoside Luteolin, 7-O-β-glucuronide Apigenin	(Rashed et al., 2015)

<i>(Maxim.) Trautv.</i> <i>(Vitaceae)</i>		
<i>Artemisia maderaspatana L.</i> <i>(Asteraceae)</i>	maderaspatana	(Jyotshna et al., 2015)
<i>Angelica decursiva Franch. &amp; Sav.</i> <i>(Apiaceae)</i>	hydroxybenzoic acid	(Yousof Ali et al., 2015)
<i>Piper bavinum C.DC.</i> <i>(Piperaceae)</i>	Bavinol A	(Dung et al., 2015)
<i>Garcinia hombroniana Pierre</i> <i>(Clusiaceae)</i>	garcihombronan N	(Jamila et al., 2015)
<i>Nelumbo nucifera Gaertn.</i> <i>(Nelumbonaceae)</i>	liensinine	(Jung et al., 2010)
<i>Evodia lepta Merr.</i> <i>(Rutaceae)</i>	kokusaginin, melineurin	(Sichaem et al., 2015)
<i>Achillea biebersteinii Hub. -Mor.</i> <i>(Asteraceae)</i>	Quercetin-7-O-β-D-glucoside, patuletin-7-O-β-D-glucoside	(Sevindik et al., 2015)
<i>Harpagophytum procumbens DC. ex Meisn.</i> <i>(Pedaliaceae)</i>	verbascosides (derivatives)	(Bae et al., 2014)
<i>Myristica fragrans Houtt.</i> <i>(Myristicaceae)</i>	(7S)-8'-(4'-hydroxy-3'-methoxyphenyl)-7-hydroxypropyl]benzene-2,4-diol, (8R,8'S )-7'-(3',4'-methylenedioxyphenyl)-8,8'-dimethyl-7-(3,4-di-hydroxyphenyl)-butane, malabaricone C	(Cuong et al., 2014)
<i>Ipomoea aquatica var. aquatica</i> <i>(Convolvulaceae)</i>	quercetin, phenolic acid, chlorogenic acid, chlorogenic acid	(Sivaraman et al., 2014)
<i>Maytenus imbricata Mart. ex Reissek</i> <i>(Celastraceae)</i>	3-oxo-11α-hydroxylup-20(29)-ene, 3-oxo-29-hydroxyfriedelane, 3,7-dioxofriedelane	(V. G. Rodrigues et al., 2014)
<i>Acanthopanax henryi Harms</i> <i>(Araliaceae)</i>	5-caffeoylequinic acid	(X. D. Zhang et al., 2014)
<i>Stemona sessilifolia</i> <i>(Friends.) Friends.</i> <i>(Stemonaceae)</i>	stenin B	(Lai et al., 2013)
<i>Amberboa ramosa</i> <i>(Roxb.) Jafri</i> <i>(Asteraceae)</i>	Amberbin	(Ibrahim et al., 2013)
<i>Murraya paniculata</i> <i>Kaneh.</i> <i>(Rutaceae)</i>	eucrestifolin	(Rehman et al., 2013)
<i>Boesenbergia rotunda</i> <i>(L.) Mansf.</i> <i>(Zingiberaceae)</i>	Boesenberginge A	(Abdelwahab, 2013)
<i>Holarrhena antidysenterica</i> <i>Wall. ex A.DC.</i> <i>(Apocynaceae)</i>	conessine, conessimine, conarrimine, conarrimine and conimine	(Z. Yang et al., 2012)
<i>Abuta grandifolia</i> <i>(Mart.) Sandwith</i> <i>(Menispermaceae)</i>	R,S)-2 N-norberbamunin, (S-S)-O4"-methyl, Nb-nor-O6'-demethyl-(+)-curine, (S-S)-O4"-methyl, O6'-demethyl-(+)-curine	(Cometa et al., 2012)
<i>Semecarpus anacardium</i> <i>Blume</i> <i>(Anacardiaceae)</i>	1',2'-dihydroxy-3'-pentadec-8-enylbenzene (A) e 1',2'-dihydroxy-3'-pentadeca-8,11-dienylbenzene (B)	(Adhami et al., 2012)
<i>Corydalis cava</i> <i>(L.) Schweigg. &amp; Körte</i> <i>(Papaveraceae)</i>	(+)-Canadalin, (+)-Canadin, (+/-)-corycavidin, (+)-bulbocapnin	(Chlebek et al., 2011)

<i>Cyrtanthus contractus N.E.Br.</i> (Amaryllidaceae)	narciprimine	(Nair et al., 2011)
<i>Buxus sempervirens L.</i> (Buxaceae)	(+)-buxabenzamidienin	(Orhan et al., 2011)
<i>Morus lhou Koidz.</i> (Moraceae)	5'-geranyl-4'-methoxy-5,7,2'-trihydroxyflavone	(J. Y. Kim et al., 2011)
<i>Hippeastrum papilio (Ravenna) Van Scheepen</i> (Amaryllidaceae)	11 $\beta$ -hydroxygalanthamine	(De Andrade et al., 2011)
<i>Alpinia officinarum</i> (Zingiberaceae)	Galangin	(Guo et al., 2010)
<i>Esenbeckia leiocarpa Engl.</i> (Rutaceae)	leptomerin	(Cardoso-Lopes et al., 2010)
<i>Ferulago campestris (Better) Grecescu</i> (Apiaceae)	siol anisoate, epielmanticin	(Dall'Acqua et al., 2010)
<i>Cleistocalyx operculatus (Roxb.) Merr. &amp; L.M. Perry</i> (Myrtaceae)	myricetin-3'-methylether 3-O- $\beta$ -d-galactopyranoside, myricetin-3',5'-dimethylether 3-O- $\beta$ -d-galactopyranoside, quercetin, kaempferol and tamarixetin.	(Min et al., 2010)
<i>Eschscholzia californica Cham.</i> (Papaveraceae)	1-(3-hydroxy-4-methoxybenzyl)-2-methyl-6,7-methylenedioxy-1,2,3,4-tetra-hydroisoquinoline, reticulin	(Cahlíková et al., 2010)
<i>Aralia cordata Thunb.</i> (Araliaceae)	17-hydroxy-ent-kaur-15-en-19-oic acid, ent-pimar-15-en-8 $\alpha$ ,19-diol	(Jung et al., 2009)
<i>Kaempferia parviflora Wall. ex Baker</i> (Zingiberaceae)	5,7-Dimethoxyflavone	(Sawasdee et al., 2009)
<i>Iris pseudopumila Tineo</i> (Iridaceae)	Isoorientin, isovitexin	(Conforti et al., 2009)
<i>Peganum nigellastrum Bunge</i> (Nitrariaceae)	vasicinone, vasicine, harmine, deoxyasicinone, deoxivasicinone, deoxivasicine, harmaline, harmol, harman, nigelastrin I, nigelastrin II	(C.-H. Wang et al., 2009)
<i>Magnolia officinalis Rehder &amp; E.H. Wilson</i> (Magnoliaceae)	4-O-methylhonokiol	(Y. K. Lee et al., 2009)
<i>Ginger rhizomes</i> (Zingiberaceae)	6-Gingerol	(Ghayur et al., 2008)
<i>Salvia sclareoides bread.</i> (Lamiaceae)	(1beta,3beta)-lup-20(29)-ene-1,3,30-triol, nepetidine, nepeticin, lupendiol, (1beta,11alpha)-dihydroxy-lup-20(29)-en-3-one , ursolic acid, sumaresinolic acid, hederagenin	(Rauter et al., 2007)
<i>Hosta plantaginea (Lamb.) Asch.</i> (Asparagaceae)	8-Demethoxy-10-O-methylhostasine	(Y.-H. Wang et al., 2007)
<i>Skimmia laureola (DC.) Siebold &amp; Zucc. ex Walp.</i> (Rutaceae)	quinoline 5-one, ribalinin, methyl isoplatodesmine	(Rahman et al., 2006)
<i>Detarium microcarpum Guill. &amp; Perr.</i> (Fabaceae)	3,4-epoxyclerodan-13E-en-15-oic acid, 5 $\alpha$ ,8 $\alpha$ (2-oxokolavénico acid), 3,4-dihydroxyclerodan-13Z-en-15-oic acid	(Cavin et al., 2006)
<i>Stephania Venosa (Blume) Spreng.</i> (Menispermaceae)	stearanine, cyclanoline, N-methyl stepholidine	(Ingkaninan et al., 2006)
<i>Sarcococca saligna Müll.Arg.</i> (Buxaceae)	2-hydroxysalignarin-E (=2'E,20S)-20-(dimethylamino)-2 $\beta$ -hydroxy-3 $\beta$ -(tigloylamino)pregn-4-ene; 1), 5,6-dihydrosarconidine (=20S)-20-(dimethylamino)-	(Atta-your-Rahman et al., 2004)

	3 $\beta$ -(methylamino)-5 $\alpha$ -pregn-16-ene; 2), salignamine (=(20S)-20-(methylamino)-3 $\beta$ -methoxypregn-5,16-diene; 3), 2-hydroxysalignamine (=(20S)-20-(dimethylamino)-2 $\beta$ -hydroxy-3 $\beta$ -methoxypregn-5,16-diene; 4), salignarin-F (=(2'E, 20S)-20-(dimethylamino)-4 $\beta$ -hydroxy-3 $\beta$ -(tigloylamino)pregn-5-ene; 5), salonin-C (=(2'E, 20S)-20-(dimethylamino)-3 $\beta$ -(tigloylamino)pregnna-4,14-diene; 6) e N-[formyl(methyl)amino]salonin-B (=(20S)-20-[formyl(methyl)amino]-3 $\beta$ -methoxypregna-5,16-diene; 7) were isolated from the MeOH extract of Sarcococca saligna, together with the six known alkaloids dictyoflebin (8), epipacisamine-D (9), saracosine (10), iso-N-formylchonemorphine (11), sarcodinine (12) and alkaloid-C (13).	
<i>Ballota limbata</i> Benth. (Lamiaceae)	diterpenoids (derivatives)	(V. U. Ahmad et al., 2004))
<i>Salvia miltiorrhiza</i> Bunge (Lamiaceae)	diterpenoids (derivatives)	(Y. Ren et al., 2004)
<i>Murraya paniculata</i> Kaneh. (Rutaceae)	murranganone, paniculatin	(Choudhary et al., 2002)
<i>Fatoua villosa</i> Nakai (Moraceae)	zeatin	(Heo et al., 2002)

Source: Author (2023)

Inhibition of the enzyme acetylcholinesterase and butyrylcholinesterase causes increased acetylcholine in the synaptic cleft, this causes AD progression to slow. AChE is responsible for the transmission of nerve impulses in vertebrates and invertebrates and hydrolysis of ACh (Tuzimski & Petruczynik, 2022). Butyrylcholinesterase has a role in cholinergic hydrolysis (ACh) and hydrolysis of organic compounds such as choline esters, butyrylcholine and succinylcholine, among others, such as the alkaloid cocaine and heroin and acetylsalicylic acid (ASA). In the study it is possible to see in (Table 2) studies d (Stefanello, 2003) the plants that have a double inhibitory action of AChE and BChE can be seen of great therapeutic interest for pharmaceutical industries and researchers, their isolated compounds are able to benefit public health and several people with Alzheimer's disease, totaling 121 documents collected in the Scopus database.

It is possible to observe the percentage of studies in relation to the botanical families and the authors, there are a total of 51 families mentioned in table 2, and using the percentage of studies dividing the number of families individually by the total document it is possible to observe that 1% of the researches are related to the families *Annonaceae*, *Asparagaceae*, *Amaranthaceae*, *Anacardiaceae*, *Combrateceae*, *Cornaceae*, *Crassulaceae*, *Caprifoliaceae*, *Convolvulaceae*, *Celastraceae*, *Dioscoreaceae*, *Iridaceae*, *Juncaceae*, *Lythraceae*, *Lauraceae*, *magnoliaceae*, *Myrtaceae*, *Nelumbonaceae*, *Nitrariaceae*, *Polygonaceae*, *Pedaliaceae*, *Rhamnaceae*, *Ranunculaceae*, *Scrophulariaceae*, *Stemonaceae*, *Vitaceae* with 1 survey each.



The researches carried out in the families Asteraceae, Apiaceae, Acanthaceae, Araliaceae, Berberidaceae, Buxaceae, Clusiaceae, Gentianaceae, Liliaceae, Malvaceae, Menispermaceae, Myristicaceae, Orchidaceae, Pterocarpaceae, Plantaginaceae, Papaveraceae and Rosaceae, totaled 2%, - 2 to 3 files each. 3% contains the families Moraceae, rubiaceae with 3 articles

*Apocynaceae, Zingiberaceae* comprise 4% with 5 documents, 6% *Lamiaceae* with 7 articles, 7% *Rutaceae* with 8 articles, and with the highest rate of appearance we have the family *Fabaceae* with 11% totaling 13 documents presented.

The *Fabaceae* was named for the first time 1789 commonly known as leguminosae is a characteristic family with the presence of flowers distributed in different geographical regions, cultivated for economic interest in the world is considered the third largest existing terrestrial family surpassed only by the families Asteraceae and Orchidaceae According to (Christenhusz & Byng, 2016) World Flora Online (2023) the families Fabaceae It has about 70,898 synonyms and 789 genera.

This family has a specific chemical particularity, for example, the isoflavonoid pterocarpans discovered in *Fabaceae*; and many of the compounds present are toxic and others are strongly found such as kaempferol, quercetin and myricetin (or myricetin). The subclass papilioideae makes up the largest subfamily of the *Fabaceae*, isoflavonoids being found in a large proportion of them (Machado et al., 2020)

For a long time the use of herbal medicines has always been used pharmacological and plant studies for treatment related to the central nervous system (CNS), has been studied and raised for example Tettevi et al (2022) prepared a review on African plants and foods related to the treatment of Alzheimer's disease listing several plants with benefits for memory, anticholinesterase and neuroprotective inhibition, in which he addressed phenotypes related to Alzheimer's and the family *Fabaceae* among others. The author in the intention of showing applications in the study of plants for Alzheimer's listed the benefits of vegetables and the possible neuroprotectors exposing extracts and compounds isolated from plants with Anti-Alzheimer's activities, boasted the species Cheng, Lin e Lane (2021) *Glycyrrhiza inflata* B. (*Fabaceae*) and its benefits for Alzheimer's in aqueous extract and its isolated compounds.

The study carried out by the authors by the plant Alhawarri et al (2023) *Cássia timoriensis* DC, *Cássia grandis* Lf both belonging to (*Fabaceae*) isolated three compounds luteolin, cinnamic acid, 4-hydroxycinnamic acid with promising activities for Alzheimer's, in addition to these plants' other important plants of the *Fabaceae* family such as *Pongamia pinnata* (L.) Pierre, *Caragana balchaschensis* (Kasn. ex Kom.) Pojark, *Castanopsis cuspidata* var. *sieboldii* (Makino) Nakai, *Astragalus membranaceus* Fisch. ex Bunge, *Acacia auriculiformis* A. CUNN Ex. Benth., *Erythrina caffra* Blanco, *Cassia obtusifolia* L., *Tephrosia purpurea* (L.) Pers., *Pueraria Lobata*, *Xylia Xylocarpa*



(Roxb.) W. Theob., *Detarium Microcarpum* Guill. & Perr. They obtained admirable results for the treatment of AD.

The pharmacological treatments currently used are substances whose action is to increase the levels of acetylcholine in the brain. These plants that act on the central nervous system in cholinergic action are expressly relevant for medicine. In particular, plants produce two types of substances called primary metabolites responsible for plant and secondary development that include essential oils (OE) (terpenes) being entirely of plant origin, these substances make available interests to pharmaceutical, cosmetic and food industries (Akman, et al 2023). These terpenes have the ability to cross biological barriers through interactivity with endogenous molecules (Ali et al., 2017).

EOs are mostly found in flowers, leaves, seeds and are usually isolated by hydrodistillation method where the extract is in direct interaction with water at the time it goes into boiling transports volatile compounds such as oil. (Ali et al., 2017) OE as examples of *ducosterol*, *alpha-spinasterol* among others presented in table 2- contains inhibition activity of the enzyme acetylcholinesterase ensuring new target molecules for the treatment of Alzheimer's. (Abbas-MOHAMMADI et al., 2022)

The therapy for Alzheimer's as well as the pathology itself has several unknowns as over the years the progression of the pathology causes numerous physiological processes such as lipid peroxidations which is a chain reaction caused by organic substances called fatty acid (hydrocarbons) that cause changes in the cell membrane influencing the permeability, fluidity and activity of the cell that can be coated by antioxidant drugs that make peroxidation impossible (BUTTERFIELD, 2020 ; YIN et al., 2016 ).

Several researchers demonstrate that the use of plants with antioxidant activity containing substances such as vitamin C, vitamin E, beta-carotene can benefit memory loss, these antioxidant compounds have the ability to neutralize free radicals inhibiting health problems. But free radicals do not only cause harm to the human being their normal production is important for cellular respiration and our survival, only their development can cause diseases to be human (LAUER et al., 2022).

The instability in antioxidant defense and increase of free radicals present in the brain produces stress oxidative which is another factor associated with Alzheimer's, as well as the occurrence of senile plaques formed by the abnormal metabolism of the amyloid beta protein, neurofibrillary tangles formed by the excess of Tau proteins that arose due to the action of hyperphosphorylation. All these events cause complications in the brain producing symptoms prevalent in various neurological pathologies (Griñán-Ferré et al., 2021).

These insertions lead to severe brain atrophy, neurodegeneration and neuroinflammation mediated by innate cells present in the brain mentioned in these authors: neuroinflammation comes from tissue damage caused by neurotoxic substances, which produces several complex chemical actions and because Alzheimer's is quite hermetic and multifactorial, the

initial state of neuroinflammation is not yet known, it is only known that Machado et al (2020) Astrocytes and microglia found near senile plaques can trigger the process of the participation of immune system compounds.

Considering the pathophysiology of the disease, biochemical research with the intention of discovering treatment, it was possible to contact in the brain variations of the enzyme MAO-A and B in high amounts, indicating that the activity of MAOs may also be the reason for AD progression. MAOs have the ability to activate beta-secretase and gamma-secretase action by amplifying the disordered generation of amyloid plaques. (Behl et al., 2021)

A study carried out by on the plant (Mahnashi et al., 2021) *Notholirion thomsnianum* (Royale) Stapf, isolating the flavone 2-(3,4-dimethoxyphenyl)-3,7-dihydroxy-4H-cromen-4-one, in its tests showed that the flavone exhibited excellent inhibition of the enzymes AChE and BChE, also contacted the inhibition of the inflammation signaling pathways COX-1 and COX-2, which may contribute to neuroinflammation and in the antioxidant test the authors denoted a relevant increase in antioxidant enzymes caused by the isolated. Similar results can be found in several studies as shown in table 3. Plant species with antioxidant, anti-inflammatory and neuroprotective activity.

Table 3. Plants with antioxidant, anti-inflammatory and neuroprotective activity in Alzheimer's disease

<b>Plants and Families</b>	<b>Anti-Alzheimer's Compounds</b>	<b>Author</b>
<i>Nardostachys jatamansi</i> (D. Don) DC. (Caprifoliaceae)	actinidine, glaziovine	(Krishnan et al., 2022)
<i>Rubus chingii</i> Hu. (Rosaceae)	ellagic acid, tiliroside and kaempferol-3-o-rutoside	(Wu et al., 2022)
<i>Prangos uechtritzii</i> Boiss. & Hausskn. (Apiaceae)	(+)-Falcarindiol and imperatorin	(Albayrak et al., 2022)
<i>Atractylodes macrocephala</i> Koidz. (Asteraceae)	Biatractylolide	(Q. Hu et al., 2022)
<i>Agathophora alopecuroides</i> (Delile) Fenzl ex Bunge, <i>Bassia indica</i> (Wight) AJ Scott (Amaranthaceae)	N-trans-feruloyl-3-methoxytyramine, N-trans-feruloyltyramine, S-(-)-3-(4-hydroxy-3-methoxyphenyl)-N-[2-(4-hydroxyphenyl)]-methoxyethyl acrylamide, N-trans-caffeoyletyramine	(Othman et al., 2022)
<i>Ocimum basilicum</i> var. <i>basilicum album</i> (L.) Benth. (Lamiaceae)	5,7-dihydroxy-3',4',5'-trimethoxyflavone and 3-hydroxy-3',4',5'-trimethoxyflavone	(Singh et al., 2022)
<i>Nelumbo nucifera</i> Gaertn. (Nelumbonaceae)	Neferine	(Tang et al., 2022)
<i>Bacopa occultans</i> (Hiern) Hutch. & Dalziel (Plantaginaceae)	bacoside-A3	(Q.-K. Bai & Zhao, 2022)
<i>Geophila repens</i> (L.) IM Johnst (Rubiaceae)	genistin, quercetin-3-D-galactoside, 9,12,15-octadecatrienoic-acid methyl-ester, phytol, retinal,	(Dash et al., 2019)

	stigmasterol, n-hexadecanoic acid, $\beta$ -sitosterol	
<i>Acorus calamus L.</i> (Acoraceae)	$\alpha$ -asarone	(Venkatesan, 2022)
<i>Rhinacanthus nasutus (L.) Kurz</i> (Acanthaceae)	Rhinacanthin-C	(Rakkhittawattana et al., 2022)
<i>Munronia henryi Harms</i> (Meliaceae)	Munronin V	(Yan et al., 2022)
<i>Myrsine seguinii H.Lév.</i> (Primulaceae)	(2R,3S)-4"-O-galloylisoastilbin, (2R,3R)-4"-O-(4"-Hydroxybenzoyl)astilbin, (2R,3R)-3"-O-E-Feruloylastilbin	(H.-J. Lee et al., 2021)
<i>Xysmalobium undulaum</i> (Apocynaceae)	acetylated glycosydated crotoxogenin, xysmalogenin-3, $\beta$ -d-glucopyranoside	(Thakur et al., 2021)
<i>Eucommia ulmoides Olive.</i> (Eucommiaceae)	ulmoidol	(Han et al., 2021)
<i>Litchi chinensis Sonn.</i> (Sapindaceae)	Jasmonates, terpenes (derivatives)	(X. Zhang et al., 2021)
<i>Silybum marianum (L.) Gaertn.</i> (Asteraceae)	silibinin A	(Esselun et al., 2021)
<i>Erythrina corallodendron var. orientalis L.</i> (Fabaceae)	10,11-Dioxo-6,7a-erythraline epoxide	(Aboelmagd et al., 2021)
<i>Murraya koenigii (L.) Spreng.</i> (Rutaceae)	9-Benzyl-9H-carbazol-4-ol	(Yano, Nakashima, Kasa, et al., 2020)
<i>Nelumbo nucifera Gaertn.</i> (Nelumbonaceae)	Asimilobine, N-methylasimilobine	(Yano, Nakashima, Oda, et al., 2020)
<i>Quercus serrata Thunb.</i> (Fabaceae)	triterpenóides (derivatives)	(May et al., 2020)
<i>Elaeagnus glabra f. oxyphylla</i> (Elaeagnaceae)	procyanidin B3, procyanidin B4 and helichryside (13)	(Y. J. Kim et al., 2020)
<i>Xanthoceras sorbifolia Bunge</i> (Sapindaceae)	barrigenol (derivatives)	(W. Li et al., 2020)
<i>Corydalis tomentella Franch.</i> (Papaveraceae)	isoquinolines	(Y.-M. Wang et al., 2020)
<i>Ceiba pentandra (L.) Gaertn.</i> (Bombacaceae)	cinchonain	(Abouelela et al., 2020)
<i>Garcinia mangostana</i> (Clusiaceae)	$\alpha$ -mangostin	(Pole et al., 2020)
<i>Zingiber officinale Roscoe</i> (Zingiberaceae)	[6]-gingerol, [8]-gingerol and [6]-shogaol	(Simon et al., 2020)
<i>Xanthoceras sorbifolium bunge</i> (Sapindaceae)	3-O-[ $\beta$ -D-glucopyranosyl (1 $\rightarrow$ 6)]-(2'-angeloyl)- $\beta$ -D-glucopyranosyl-28-O- $\beta$ -D-glucopyranosyl(1 $\rightarrow$ 6)[ $\alpha$ -L-rhamnopyranosyl(1 $\rightarrow$ 2)- $\beta$ -D-glucopyranosyl]-21-O-acetyl-16-deoxybarringtogenol C	(Thing et al., 2020)
<i>Isodon japonicus (Burm.f.) H.Hara</i> (Lamiaceae)	Rosmarinic Acid	(Sun et al., 2019)
<i>Abronia nana S.Watson</i> (Nyctaginaceae)	boeravinone x	(E.-J. Yang et al., 2019)
<i>Lawsonia inermis L.</i> (Lythraceae)	1,2,4-Trihydroxynaphthalene-2-O- $\beta$ -D-glucopyranoside	(Dhouafli et al., 2019)
<i>Sophora tonkinensis</i> (Fabaceae)	sophotokin	(Xia et al., 2019)
<i>Cirsium maackii Maxim.</i> (Asteraceae)	luteolin	(Wagle et al., 2019)

<i>Schinus polygamus</i> var. <i>chilensis</i> F.A. Barkley (Anacardiaceae)	Agathisflavone	(Dumitru et al., 2019)
<i>Lycoris ×chejuensis</i> Kurita & P.S. Hsu (Amaryllidaceae)	7-deoxy-trans-dihydronarciclasine	(D. Zhao et al., 2019)
<i>Pithecellobium Clypearia</i> (Jack) Benth. (Fabaceae)	(2 R ,3 R)-7,8,3 ',4'-tetrahidroxidihydro-flavonol	(Y.-X. Wang et al., 2017)
<i>Reynoutria sachalinensis</i> Nakai (Polygonaceae)	1-Decanol, campesterol, ergosterol peroxide, quercetin and isoquercitrin	( Eom et al., 2017)
<i>Pyrola decorate</i> H. Andr (Pyrolaceae)	Betulin, ursolic acid , monotropein	(X. Yang et al., 2017)
<i>Angelica gigas</i> var. <i>minor</i> Momiy. (Apiaceae) <i>Scutellaria baicalensis</i> Georgi (Lamiaceae)	decursin, Wogonin	(H. W. Lee et al., 2017)
<i>Aframomum melegueta</i> var. <i>violaceum</i> (Ridl.) K. Schum. (Zingiberaceae)	Gingerol	(El Halawany et al., 2017)
<i>Monsonia angustifolia</i> E. Mey. (Geraniaceae)	justicidin A	(Chun et al., 2017)
<i>Akebia quinata</i> (Thunb. ex Houtt.) Decne. (Lardizabalaceae)	akequintoside F, collinsonidin, akebonic acid, hederagenin, asperosaponin C	(Chowdhury et al., 2017)
<i>Pteris multifida</i> Poir (Pteridaceae)	2b,15α-dihydroxy-ent-kaur-16-ene, pterokaurane P1	(J. W. Kim et al., 2017)
<i>Fumaria officinalis</i> (Papaveracea)	Parfumidine, sinactine	(Chlebek, Novák, et al., 2016)
<i>Sophora flavescens</i> Aiton (Fabaceae)	maackiain	(H. W. Lee et al., 2016)
<i>Siegesbeckia pubescens</i> (Asteraceae)	5,3'-dihydroxy-3,7,4'-trimethoxyflavone	(D.-S. Lee et al., 2016)
<i>Valeriana amurensis</i> P.A. Smirn. ex Kom. (Caprifoliaceae)	(+)-medioresinol-4,4'-di-O-β-D-glucopyranoside, (+)-syringaresinol-4,4'-di-O-β-D-glucopyranoside, prinsepiol-4-O-β-D-glucopyranoside , (+)-8,8'-dihydroxy-pinoresinol-4,4'-di-O-β-D-glucopyranoside, prinsepiol, and 6 iridoids of jatamanin A, 7-hydroxy-8-(hydroxymethyl)-4-methylenehexahydrocyclopenta[c] pyran-1(3H)-one , 4-hydroxymethyl-cyclopenta[c] pyran-7-carboxaldehyde , patriscabroside III , jatamanin E, patrinoside .	(Wan et al., 2016)
<i>Sophora flavescens</i> Aiton (Fabaceae)	Maackiain	(H. W. Lee et al., 2016)
<i>Uncaria rhynchophylla</i> Miq. (Rubiaceae)	Uncarinic Acid C	(Yoshioka et al., 2016)
<i>Prunus persica</i> (L.) Batsch (Rosaceae)	methyl amygdalinate	(X. Zhao et al., 2016)
<i>Corydalis cava</i> (L.) Schweigg. & Körte (Papaveracea)	(-) -corycavamine, (+) -corynoline	(Chlebek, De Simone, et al., 2016)
<i>Corydalis cava</i> (Fumariacea)	(-) -corycavamine, (+) -corynoline	(Chlebek et al., 2011)
<i>Zelkova serrata</i> Makino (Ulmacea)	7-Hydroxycalamenene	(Yen et al., 2016)

<i>Lycium barbarum</i> Mill. (solanaceae)	dicaffeoylspermidine (derivatives)	(Zhou et al., 2016)
<i>Garcinia mangostana</i> L.(Clusiaceae)	$\alpha$ -Mangostin, Gartanin, Garinone C, C-Mangostin	(S.-N. Wang et al., 2016)
<i>Serjania erecta</i> Radlk. (Sapindaceae)	vitexin	(Guimarães et al., 2015)
<i>Angelica shikokiana</i> Makino ex Y. Yabe (Apiaceae)	quercetin, hyuganin E, isoepoxyterixin.	(Mira et al., 2015)
<i>Vitis thunbergii</i> var. <i>taiwaniana</i> F.Y. Lu (Vitaceae)	miyabenol C	(J. Hu et al., 2015)
<i>Acer nikoense</i> (Miq.) Maxim. (Sapindaceae)	Acerogenin A	(D.-S. Lee et al., 2015)
<i>Origanum glandulosum</i> Desf. (Lamiaceae)	Rosmarinic acid, globoidnan A	(Bash et al., 2014)
<i>Disporum viridescens</i> (Maxim.) Nakai (Colchicaceae)	(+)-dihydrodehydrodiconiferyl alcohol-9-O- $\beta$ -D- glucopyranoside, (-)-9'- hydroxypinoresinol	(Cho et al., 2014)
<i>Jatropha multifida</i> L. (Euphorbiaceae)	Apocynin	(T Hart et al., 2014)
<i>Torreya yunnanensis</i> WCCheng & LKFu (Taxaceae)	( $\pm$ ) -Torreyunlignans (derivatives)	(Z.-B. Cheng et al., 2014)
<i>Celastrus orbiculatus</i> Thunb. (Celastraceae)	(M)-bicelaphanol A	(X. J. Wang et al., 2013)
<i>Orobanche minor</i> Sm. (Orobanchaceae)	acteoside (1a)	(Kurisu et al., 2013)
<i>Lycoris radiata</i> (L'Hér.) Herb. (Amaryllidaceae)	1,2-Di-O-acetyllycorine, 1-O- acetyllycorine	(Xin et al., 2013)
<i>Perilla frutescens</i> (L.) Britton (Lamiaceae)	Luteolin	(G. Zhao et al., 2012)
<i>Uncaria rhynchophylla</i> Miq. (Rubiacae)	rhynchophylline, isorhynchophylline	(Xian et al., 2012)
<i>Valeriana amurensis</i> P.A. Smirn. ex Kom. (Caprifoliaceae)	sesquiterpenoids (derivatives) lignans(derivatives)	(Q. Wang et al., 2012)
<i>Ginkgo biloba</i> L. (Ginkgoaceae)	Isorhamnetin	(Xu et al., 2012)
<i>Itoa orientalis</i> Hemsl. (Salicaceae)	Xylocoside G	(Yu et al., 2012)
<i>Erigeron annuus</i> Sessé & Moc. (Asteraceae)	Caffeic acid	(Jeong et al., 2011)
<i>Eleutherococcus</i> <i>senticosus</i> Maxim. (Araliaceae)	eleutheroside B, eleutheroside E, isofraxidin	(Y. Bai et al., 2011; Jeong et al., 2011)
<i>Euclea crispa</i> subsp. <i>Crispa</i> (Ebenaceae) <i>Crinum macowanii</i> Baker (Amaryllidaceae)	3-oxo-oleanolic acid, natalenone, lycorine, hamayne	(Kwon et al., 2011)
<i>Iris tenuifolia</i> Pall. (Iridaceae)	3'-hydroxy-5,7-dimethoxy-4-O- 2'-cycloflavan, 3'-hydroxy-5- methoxy-6,7- methylenedioxy- 4-O-2'-cycloflavan	(Cui et al., 2011)
<i>Siegesbeckia glabrescens</i> (Asteraceae)	3, 4'-dimethylquercetin, 3, 7- dimethylquercetin, 3- methylquercetin and 3, 7, 4'- trimethylquercetin	(Lim et al., 2011)
<i>Pueraria lobata</i> (Willd.) Ohwi (Fabaceae)	genistein, biochanin A	(Choi et al., 2010)

<i>Rhodiola rosea L.</i> (Crassulaceae)	salidroside	(Na et al., 2010; L. Zhang et al., 2010)
<i>Eragrostis ferruginea P. Beauv.</i> (Poaceae)	Tricin	(Na et al., 2010)
<i>Hypericum perforatum L.</i> (Hypericaceae)	hyperforin	(Denmark et al., 2006)
<i>Uncaria rhynchophylla</i> (Miq.) Jacks. (Rubiaceae)	(+) -catechin, (-) -epicatechin	(Hou et al., 2005)
<i>Flemingia macrophylla</i> (Willd.) Kuntze ex Merr. (Fabaceae)	flemingichromone, osajin, 5,7,4'-trihydroxy-6,8'- diprenylisoflavone, 5,7,4'- trihydroxy-6,3'- diprenylisoflavone, aureole	(Shiao et al., 2005)
<i>Turmeric longa L.</i> (Zingiberaceae)	calebin-A, curcumin, bisdemethoxycurcumin, 1,7- bis(4-hydroxyphenyl)-1- heptene-3,5-dione	(S.-Y. Park & Kim, 2002)

Table 3 shows the total of 50 botanical families. Applying the percentage, it is found that 1% of the families have anti-Alzheimer's activity, such as Acoraceae, Acanthaceae, Apocynaceae, Araliaceae, Anacardiaceae, Bombacaceae, Brassicaceae, Celastraceae, Colchicaceae, Crassulaceae, Eucammiaceae, Elaeagnaceae, Euphorbiaceae, Ebenaceae, Fumariaceae, Ginkgoaceae, Hypericaceae, Iridaceae, Lythraceae, Lardizabalaceae, Meliaceae, Nyctaginaceae, Orobanchaceae, Polygonaceae, Pyrolaceae, Pteridaceae, Plantaginaceae, Primulaceae, Poaceae, Rutaceae, Solanaceae, Salicaceae, Taxaceae and Ulmaceae, each with 1 document.

The families Clusiaceae, Geramiaceae, Nelumbonaceae and Rosaceae represent 2% of the plants mentioned in table 3, each containing 2 documents.

Apiaceae, Amaryllidaceae, Caprifoliaceae, Papaveraceae and Zingiberaceae exhibit 3% of the studies collected. 4% correspond only to the Rubiaceae family, which presented 4 articles. 6% correspond to the Lamiaceae family. The family Sapindaceae has 5 studies, representing 5% of the studies. 7% correspond to the Asteraceae family, with 6 published articles.

The most studied family, shown in the table, was the Fabaceae, with 11% of the research and 10 documents presented.

The medicinal plants mentioned in Table 3 in turn presented by families, species, isolated compound responsible for the pharmacological activity include antioxidant mechanism in the direct elimination of reactive oxygen species (ROS), in the activation of enzymes that have antioxidant activity, activity of metal chelating agent, reinforcement of the amount of  $\alpha$ -tocopherol radicals, blocking of NAPDH, relief of oxidative stress caused by nitric oxide, addition of uric acid levels and increase of substances with antioxidant characteristics and antioxidant molecules of low molecular weight (Kurutas, 2016)

The antioxidant activity may be present in several chemical compounds isolated from plants, called natural antioxidant, used in several groups of secondary metabolites, such as flavonoids (catechins) and tannins (proanthocyanidins), among others. Catechins are formidable antioxidants



thanks to the group of hydroxyls present in their molecules, the elimination of free radicals present in the body can also be generated by molecules that contain double bond in the phenolic ring, hydroxyl side chain molecules, anthocyanidin glycosylation among others, can also be classified as synthetic antioxidant widely used in the food industry (Collins et al., 2022)

The use of plant antioxidants has many contradictions by some researchers plus studies that point out that antioxidant supplementation by external means is one of the most favorable methods in progression in the development of therapies for neurological diseases caused by radicals' books. (Kasote et al., 2015)

## 5 CONCLUSIONS

Based on the analysis carried out, it can be concluded that there has been a significant development in the number of publications of studies related to plants, isolated compounds and Alzheimer's diseases over the years. Although there have been fluctuations, especially in the period of the COVID-19 pandemic, the number of publications has increased substantially, indicating a growing interest in this field of research.

The most productive author identified was Choi, JS, who contributed 17 papers addressing studies of plants, Alzheimer's disease, cholinergic disease, and neuroprotection. China and South Korea stood out as the main producers of articles on the subject, with a significant number of publications.

The high incidence of Alzheimer's disease in China may be a factor driving interest and research in this area. The rapid aging of the Chinese population and the increase in age-related problems, including appearance, pose challenges to public health and society as a whole. The costs associated with treating Alzheimer's disease are also long-lasting and likely to increase in the future.

The analysis of the advanced and themes revealed the importance of the keywords "Alzheimer's disease" and "plant extract" in the studies carried out. The collaboration between the researcher, evidenced by the network of co-occurrence of the keywords, demonstrates the construction of knowledge over time.

Studies on medicinal plants that inhibit the enzymes acetylcholinesterase (AChE) and butyrylcholinesterase (BuChE) have been highlighted as relevant to the development of drugs in the treatment of Alzheimer's disease. Methods such as Ellman's technique and thin layer chromatography were used to detect enzymatic inhibitory activity and track plant extracts.

In summary, productivity analysis has revealed a growing interest in research on plants, isolated compounds, and Alzheimer's disease. These studies aim to find therapeutic solutions for Alzheimer's disease, which pose a significant public health challenge in several countries, including China. The



development of new medicines based on medicinal plants and the search for compounds that inhibit disease-related enzymes are promising areas of research.



## REFERENCES

- Abbas-Mohammadi, M., Moridi Farimani, M., Salehi, P., Ebrahimi, S. N., Sonboli, A., Kelso, C., & Skropeta, D. (2022a). Molecular networking based dereplication of AChE inhibitory compounds from the medicinal plant *Vincetoxicum funebre* (Boiss. & Kotschy). *Journal of Biomolecular Structure and Dynamics*, 40(5), 1942–1951. <https://doi.org/10.1080/07391102.2020.1834455>
- Abdelwahab, S. I. (2013). In vitro inhibitory effect of Boeserngin A on human acetylcholinesterase: Understanding its potential using in silico ADMET studies. *Journal of Applied Pharmaceutical Science*, 3(3), 30–35. <https://doi.org/10.7324/JAPS.2013.30306>
- Aboelmagd, M., Elokely, K. M., Said, A., Haggag, E. G., Ghoneim, M. M., & Ross, S. A. (2021). New selective human mao-b inhibitors from the stems of erythrina corallodendron l. *Records of Natural Products*, 15(5), 368–379. <https://doi.org/10.25135/rnp.229.21.01.1940>
- Abouelela, M. E., Orabi, M. A. A., Abdelhamid, R. A., Abdelkader, M. S. A., Darwish, F. M. M., Hotsumi, M., & Konno, H. (2020). Anti-Alzheimer's flavanolignans from Ceiba pentandra aerial parts. *Fitoterapia*, 143. <https://doi.org/10.1016/j.fitote.2020.104541>
- Adhami, H. R., Linder, T., Kaehlig, H., Schuster, D., Zehl, M., & Krenn, L. (2012). Catechol alkenyls from Semecarpus anacardium: Acetylcholinesterase inhibition and binding mode predictions. *Journal of Ethnopharmacology*, 139(1), 142–148. <https://doi.org/10.1016/j.jep.2011.10.032>
- Ahammed, S., Afrin, R., Uddin, N., Al-Amin, Y., Hasan, K., Haque, U., Islam, K. M. M., Alam, A. H. M. K., Tanaka, T., & Sadik, G. (2021). Acetylcholinesterase inhibitory and antioxidant activity of the compounds isolated from Vanda roxburghii. *Advances in Pharmacological and Pharmaceutical Sciences*, 2021. <https://doi.org/10.1155/2021/5569054>
- Ahmad, H., Ahmad, S., Ali, M., Latif, A., Shah, S. A. A., Naz, H., ur Rahman, N., Shaheen, F., Wadood, A., Khan, H. U., & Ahmad, M. (2018). Norditerpenoid alkaloids of Delphinium denudatum as cholinesterase inhibitors. *Bioorganic Chemistry*, 78, 427–435. <https://doi.org/10.1016/j.bioorg.2018.04.008>
- Ahmad, V. U., Farooq, U., Abbaskhan, A., Hussain, J., Abbasi, M. A., Nawaz, S. A., & Choudhary, M. I. (2004). Four New Diterpenoids from Ballota limbata. *Helvetica Chimica Acta*, 87(3), 682–689. <https://doi.org/10.1002/hlca.200490064>
- Ajayi, O. S., Aderogba, M. A., Obuotor, E. M., & Majinda, R. R. T. (2019). Acetylcholinesterase inhibitor from Anthocleista vogelii leaf extracts. *Journal of Ethnopharmacology*, 231, 503–506. <https://doi.org/10.1016/j.jep.2018.11.009>
- Akram, M., & Nawaz, A. (2017). Effects of medicinal plants on alzheimer's disease and memory deficits. Em *Neural Regeneration Research* (Vol. 12, Número 4, p. 660–670). Medknow Publications. <https://doi.org/10.4103/1673-5374.205108>
- Al Mamun, A., Maříková, J., Hulcová, D., Janoušek, J., Šafratová, M., Nováková, L., Kučera, T., Hrabinová, M., Kuneš, J., Korábečný, J., & Cahlíková, L. (2020). Amaryllidaceae alkaloids of belladine-type from narcissus pseudonarcissus cv. Carlton as new selective inhibitors of butyrylcholinesterase. *Biomolecules*, 10(5). <https://doi.org/10.3390/biom10050800>
- Albayrak, G., Demir, S., Koyu, H., & Baykan, S. (2022). Anticholinesterase Compounds from Endemic Prangos uechtritzii. *Chemistry and Biodiversity*, 19(11). <https://doi.org/10.1002/cbdv.202200557>



Alhawarri, M. B., Dianita, R., Rawa, M. S. A., Nogawa, T., & Wahab, H. A. (2023). Potential Anti-Cholinesterase Activity of Bioactive Compounds Extracted from Cassia grandis L.f. and Cassia timoriensis DC. *Plants*, 12(2). <https://doi.org/10.3390/plants12020344>

Ali, M., Muhammad, S., Shah, M. R., Khan, A., Rashid, U., Farooq, U., Ullah, F., Sadiq, A., Ayaz, M., Ali, M., Ahmad, M., & Latif, A. (2017). Neurologically potent molecules from Crataegus oxyacantha; isolation, anticholinesterase inhibition, and molecular docking. *Frontiers in Pharmacology*, 8(JUN). <https://doi.org/10.3389/fphar.2017.00327>

Ali, R., Atia-tul-Wahab, Wajid, S., Khan, M. A., Yousuf, S., Shaikh, M., Hassan Laghari, G., Rahman, A.-U., & Choudhary, M. I. (2022). Isolation, derivatization, in-vitro, and in-silico studies of potent butyrylcholinesterase inhibitors from Berberis parkeriana Schneid. *Bioorganic Chemistry*, 127. <https://doi.org/10.1016/j.bioorg.2022.105944>

Aria, M., & Cuccurullo, C. (2017). bibliometrix : An R-tool for comprehensive science mapping analysis. *Journal of Informetrics*, 11(4), 959–975. <https://doi.org/10.1016/j.joi.2017.08.007>

Atta-Ur-Rahman, Zaheer-Ul-Haq, Feroz, F., Khalid, A., Nawaz, S. A., Khan, M. R., & Choudhary, M. I. (2004). New Cholinesterase-Inhibiting Steroidal Alkaloids from Sarcococca saligna. *Helvetica Chimica Acta*, 87(2), 439–448. <https://doi.org/10.1002/hlca.200490042>

Ayaz, M., Sadiq, A., Junaid, M., Ullah, F., Subhan, F., & Ahmed, J. (2017). Neuroprotective and anti-aging potentials of essential oils from aromatic and medicinal plants. Em *Frontiers in Aging Neuroscience* (Vol. 9, Número MAY). Frontiers Research Foundation. <https://doi.org/10.3389/fnagi.2017.00168>

Bae, Y. H., Cuong, T. D., Hung, T. M., Kim, J. A., Woo, M. H., Byeon, J. S., Choi, J. S., & Min, B. S. (2014). Cholinesterase inhibitors from the roots of Harpagophytum procumbens. *Archives of Pharmacal Research*, 37(9), 1124–1129. <https://doi.org/10.1007/s12272-013-0316-y>

Bai, Q.-K., & Zhao, Z.-G. (2022). Isolation and neuronal apoptosis inhibitory property of bacoside-A3 via downregulation of  $\beta$ -amyloid induced inflammatory response. *Biotechnology and Applied Biochemistry*, 69(2), 726–734. <https://doi.org/10.1002/bab.2147>

Bai, Y., Tohda, C., Zhu, S., Hattori, M., & Komatsu, K. (2011). Active components from Siberian ginseng (*Eleutherococcus senticosus*) for protection of amyloid  $\beta$ (25-35)-induced neuritic atrophy in cultured rat cortical neurons. *Journal of Natural Medicines*, 65(3–4), 417–423. <https://doi.org/10.1007/s11418-011-0509-y>

Balaei-Kahnamoei, M., Saeedi, M., Rastegari, A., Shams Ardekani, M. R., Akbarzadeh, T., & Khanavi, M. (2021). Phytochemical Analysis and Evaluation of Biological Activity of Lawsonia inermis Seeds Related to Alzheimer's Disease. *Evidence-Based Complementary and Alternative Medicine*, 2021. <https://doi.org/10.1155/2021/5965061>

Basli, A., Delaunay, J.-C., Pedrot, E., Bernillon, S., Madani, K., Monti, J.-P., Mérillon, J.-M., Chibane, M., & Richard, T. (2014). New cyclolignans from Origanum glandulosum active against  $\beta$ -amyloid aggregation. *Records of Natural Products*, 8(3), 208–216. <https://www.scopus.com/inward/record.uri?eid=2-s2.0-84899754508&partnerID=40&md5=2cf11cc33e627211c15e1fc7cd7d39e9>

Behl, T., Kaur, D., Sehgal, A., Singh, S., Sharma, N., Zengin, G., Andronie-Cioara, F. L., Toma, M. M., Bungau, S., & Bumbu, A. G. (2021). Role of Monoamine Oxidase Activity in Alzheimer's Disease:



An Insight into the Therapeutic Potential of Inhibitors. *Molecules*, 26(12), 3724. <https://doi.org/10.3390/molecules26123724>

Bhakta, H. K., Park, C. H., Yokozawa, T., Tanaka, T., Jung, H. A., & Choi, J. S. (2017). Potential anti-cholinesterase and  $\beta$ -site amyloid precursor protein cleaving enzyme 1 inhibitory activities of cornuside and gallotannins from *Cornus officinalis* fruits. *Archives of Pharmacal Research*, 40(7), 836–853. <https://doi.org/10.1007/s12272-017-0924-z>

Boudjada, A., Touil, A., Bensouici, C., Bendif, H., & Rhouati, S. (2019). Phenanthrene and dihydrophenanthrene derivatives from *Dioscorea communis* with anticholinesterase, and antioxidant activities. *Natural Product Research*, 33(22), 3278–3282. <https://doi.org/10.1080/14786419.2018.1468328>

Breijeh, Z., & Karaman, R. (2020). Comprehensive Review on Alzheimer's Disease: Causes and Treatment. *Em Molecules* (Vol. 25, Número 24). MDPI. <https://doi.org/10.3390/MOLECULES25245789>

Butterfield, D. A. (2020). Brain lipid peroxidation and alzheimer disease: Synergy between the Butterfield and Mattson laboratories. *Ageing Research Reviews*, 64, 101049. <https://doi.org/10.1016/j.arr.2020.101049>

Cahlíková, L., Macáková, K., Kuneš, J., Kurfürst, M., Opletal, L., Cvačka, J., Chlebek, J., & Blunden, G. (2010). Acetylcholinesterase and butyrylcholinesterase inhibitory compounds from *Eschscholzia californica* (Papaveraceae). *Natural Product Communications*, 5(7), 1035–1038. <https://doi.org/10.1177/1934578x1000500710>

Cardoso-Lopes, E. M., Maier, J. A., Da Silva, M. R., Regasini, L. O., Simote, S. Y., Lopes, N. P., Pirani, J. R., Da Silva Bolzani, V., & Young, M. C. M. (2010). Alkaloids from stems of *esenbeckia leiocarpa* Engl. (Rutaceae) as potential treatment for alzheimer disease. *Molecules*, 15(12), 9205–9213. <https://doi.org/10.3390/molecules15129205>

Castro e Silva, J. H., Ferreira, R. S., Pereira, E. P., Braga-De-Souza, S., Alves de Almeida, M. M., dos Santos, C. C., Butt, A. M., Caiazzo, E., Capasso, R., Amaral da Silva, V. D., & Costa, S. L. (2020). Amburana cearensis: Pharmacological and neuroprotective effects of its compounds. *Molecules*, 25(15). <https://doi.org/10.3390/molecules25153394>

Cavin, A.-L., Hay, A.-E., Marston, A., Stoeckli-Evans, H., Scopelliti, R., Diallo, D., & Hostettmann, K. (2006). Bioactive diterpenes from the fruits of *Detarium microcarpum*. *Journal of Natural Products*, 69(5), 768–773. <https://doi.org/10.1021/np058123q>

Chen, H.-W., He, X.-H., Yuan, R., Wei, B.-J., Chen, Z., Dong, J.-X., & Wang, J. (2016). Sesquiterpenes and a monoterpenoid with acetylcholinesterase (AchE) inhibitory activity from *Valeriana officinalis* var. *latifolia* in vitro and in vivo. *Fitoterapia*, 110, 142–149. <https://doi.org/10.1016/j.fitote.2016.03.011>

Chen, S.-L., Yu, H., Luo, H.-M., Wu, Q., Li, C.-F., & Steinmetz, A. (2016). Conservation and sustainable use of medicinal plants: problems, progress, and prospects. *Chinese Medicine*, 11(1), 37. <https://doi.org/10.1186/s13020-016-0108-7>

Cheng, Y.-J., Lin, C.-H., & Lane, H.-Y. (2021). Involvement of Cholinergic, Adrenergic, and Glutamatergic Network Modulation with Cognitive Dysfunction in Alzheimer's Disease. *International Journal of Molecular Sciences*, 22(5), 2283. <https://doi.org/10.3390/ijms22052283>



Cheng, Z.-B., Lu, X., Bao, J.-M., Han, Q.-H., Dong, Z., Tang, G.-H., Gan, L.-S., Luo, H.-B., & Yin, S. (2014). ( $\pm$ )-torreyunlignans A-D, Rare 8-9' linked neolignan enantiomers as phosphodiesterase-9A inhibitors from *torreya yunnanensis*. *Journal of Natural Products*, 77(12), 2651–2657. <https://doi.org/10.1021/np500528u>

Chlebek, J., De Simone, A., Hošálková, A., Opletal, L., Pérez, C., Pérez, D. I., Havlíková, L., Cahliková, L., & Andrisano, V. (2016). Application of BACE1 immobilized enzyme reactor for the characterization of multifunctional alkaloids from *Corydalis cava* (Fumariaceae) as Alzheimer's disease targets. *Fitoterapia*, 109, 241–247. <https://doi.org/10.1016/j.fitote.2016.01.008>

Chlebek, J., Macáková, K., Cahliková, L., Kurfürst, M., Kuneš, J., & Opletal, L. (2011). Acetylcholinesterase and butyrylcholinesterase inhibitory compounds from *Corydalis cava* (Fumariaceae). *Natural Product Communications*, 6(5), 607–610. <https://doi.org/10.1177/1934578x1100600507>

Chlebek, J., Novák, Z., Kassemová, D., Šafratová, M., Kostelník, J., Malý, L., Ločárek, M., Opletal, L., Hošt' Álková, A., Hrabinová, M., Kuneš, J., Novotná, P., Urbanová, M., Nováková, L., Macáková, K., Hulcová, D., Solich, P., Pérez Martín, C., Jun, D., & Cahliková, L. (2016). Isoquinoline Alkaloids from *Fumaria officinalis* L. and Their Biological Activities Related to Alzheimer's Disease. *Chemistry and Biodiversity*, 13(1), 91–99. <https://doi.org/10.1002/cbdv.201500033>

Cho, N., Yang, H., Kim, J. W., Kim, Y. C., & Sung, S. H. (2014). Chemical constituents isolated from *Disporum viridescens* leaves and their inhibitory effect on nitric oxide production in BV2 microglial cells. *Bioorganic and Medicinal Chemistry Letters*, 24(24), 5675–5678. <https://doi.org/10.1016/j.bmcl.2014.10.068>

Choi, Y.-H., Hong, S. S., Shin, Y. S., Hwang, B. Y., Park, S.-Y., & Lee, D. (2010). Phenolic compounds from *Pueraria lobata* protect PC12 cells against A $\beta$ -induced toxicity. *Archives of Pharmacal Research*, 33(10), 1651–1654. <https://doi.org/10.1007/s12272-010-1014-7>

Choudhary, M. I., Azizuddin, Khalid, A., Sultani, S. Z., & Atta-ur-Rahman. (2002). A new coumarin from *Murraya paniculata*. *Planta Medica*, 68(1), 81–83. <https://doi.org/10.1055/s-2002-19874>

Chowdhury, M. A., Ko, H. J., Lee, H., Aminul Haque, M., Park, I.-S., Lee, D.-S., & Woo, E.-R. (2017). Oleanane triterpenoids from *Akebiae Caulis* exhibit inhibitory effects on A $\beta$ 42 induced fibrillogenesis. *Archives of Pharmacal Research*, 40(3), 318–327. <https://doi.org/10.1007/s12272-016-0885-7>

Christenhusz, M. J. M., & Byng, J. W. (2016). The number of known plants species in the world and its annual increase. Em *Phytotaxa* (Vol. 261, Número 3, p. 201–217). Magnolia Press. <https://doi.org/10.11646/phytotaxa.261.3.1>

Chun, Y. S., Kim, J., Chung, S., Khorombi, E., Naidoo, D., Nthambeleni, R., Harding, N., Maharaj, V., Fouche, G., & Yang, H. O. (2017). Protective Roles of *Monsonia angustifolia* and Its Active Compounds in Experimental Models of Alzheimer's Disease. *Journal of Agricultural and Food Chemistry*, 65(15), 3133–3140. <https://doi.org/10.1021/acs.jafc.6b04451>

Collins, A. E., Saleh, T. M., & Kalisch, B. E. (2022). Naturally Occurring Antioxidant Therapy in Alzheimer's Disease. Em *Antioxidants* (Vol. 11, Número 2). MDPI. <https://doi.org/10.3390/antiox11020213>

Cometa, M. F., Fortuna, S., Palazzino, G., Volpe, M. T., Rengifo Salgado, E., Nicoletti, M., & Tomassini, L. (2012). New cholinesterase inhibiting bisbenzylisoquinoline alkaloids from *Abuta grandifolia*. *Fitoterapia*, 83(3), 476–480. <https://doi.org/10.1016/j.fitote.2011.12.015>



Conforti, F., Rigano, D., Menichini, F., Loizzo, M. R., & Senatore, F. (2009). Protection against neurodegenerative diseases of Iris pseudopumila extracts and their constituents. *Fitoterapia*, 80(1), 62–67. <https://doi.org/10.1016/j.fitote.2008.10.005>

Cui, Y.-M., Wang, H., Liu, Q.-R., Han, M., Lu, Y., & Zhao, C.-Q. (2011). Flavans from Iris tenuifolia and their effects on  $\beta$ -amyloid aggregation and neural stem cells proliferation in vitro. *Bioorganic and Medicinal Chemistry Letters*, 21(15), 4400–4403. <https://doi.org/10.1016/j.bmcl.2011.06.039>

Cuong, D., Hung, T. M., Han, H. Y., Roh, H. S., Seok, J.-H., Lee, J. K., Jeong, J. Y., Choi, J. S., Kim, J. A., & Min, B. S. (2014). Potent acetylcholinesterase inhibitory compounds from Myristica fragrans. *Natural Product Communications*, 9(4), 499–502. <https://doi.org/10.1177/1934578x1400900418>

Dall'Acqua, S., Maggi, F., Minesso, P., Salvagno, M., Papa, F., Vittori, S., & Innocenti, G. (2010). Identification of non-alkaloid acetylcholinesterase inhibitors from Ferulago campestris (Besser) Grecescu (Apiaceae). *Fitoterapia*, 81(8), 1208–1212. <https://doi.org/10.1016/j.fitote.2010.08.003>

Dash, U. C., Kanhar, S., Dixit, A., Dandapat, J., & Sahoo, A. K. (2019). Isolation, identification, and quantification of Pentylcucumene from Geophila repens: A new class of cholinesterase inhibitor for Alzheimer's disease. *Bioorganic Chemistry*, 88. <https://doi.org/10.1016/j.bioorg.2019.102947>

De Andrade, J. P., Berkov, S., Viladomat, F., Codina, C., Zuanazzi, J. A. S., & Bastida, J. (2011). Alkaloids from hippeastrum papilio. *Molecules*, 16(8), 7097–7104. <https://doi.org/10.3390/molecules16087097>

Demmak, R. G., Bordage, S., Bensegueni, A., Boutaghane, N., Hennebelle, T., Mokrani, E. H., & Sahpaz, S. (2019). Chemical constituents from solenostemma argel and their cholinesterase inhibitory activity. *Natural Product Sciences*, 25(2), 115–121. <https://doi.org/10.20307/nps.2019.25.2.115>

Dhouaifi, Z., Ben Jannet, H., Mahjoub, B., Leri, M., Guillard, J., Saidani Tounsi, M., Stefani, M., & Hayouni, E. A. (2019). 1,2,4-trihydroxynaphthalene-2-O- $\beta$ -D-glucopyranoside: A new powerful antioxidant and inhibitor of A $\beta$ 42 aggregation isolated from the leaves of Lawsonia inermis. *Natural Product Research*, 33(10), 1406–1414. <https://doi.org/10.1080/14786419.2017.1419229>

Dinamarca, M. C., Cerpa, W., Garrido, J., Hancke, J. L., & Inestrosa, N. C. (2006). Hyperforin prevents  $\beta$ -amyloid neurotoxicity and spatial memory impairments by disaggregation of Alzheimer's amyloid- $\beta$ -deposits. *Molecular Psychiatry*, 11(11), 1032–1048. <https://doi.org/10.1038/sj.mp.4001866>

Ding, K., Guo, S., Rong, W., Li, Q., Liu, R., Xu, H., Yin, Y., & Bi, K. (2020). A new oleanane type pentacyclic triterpenoid saponin from the husks of xanthoceras sorbifolium bunge and its neuroprotection on PC12 cells injury induced by A $\beta$ 25-35. *Natural Product Research*, 34(22), 3212–3218. <https://doi.org/10.1080/14786419.2018.1557172>

Dumitru, G., El-Nashar, H. A. S., Mostafa, N. M., Eldahshan, O. A., Boiangiu, R. S., Todirascu-Ciornea, E., Hritcu, L., & Singab, A. N. B. (2019). Agathisflavone isolated from Schinus polygamus (Cav.) Cabrera leaves prevent scopolamine-induced memory impairment and brain oxidative stress in zebrafish (*Danio rerio*). *Phytomedicine*, 58. <https://doi.org/10.1016/j.phymed.2019.152889>

Dung, H. V., Cuong, T. D., Chinh, N. M., Quyen, D., Kim, J. A., Byeon, J. S., Woo, M. H., Choi, J. S., & Min, B. S. (2015). Compounds from the aerial parts of Piper bavinum and their anti-cholinesterase activity. *Archives of Pharmacal Research*, 38(5), 677–682. <https://doi.org/10.1007/s12272-014-0432-3>



Dzoyem, J. P., Nkuete, A. H. L., Ngameni, B., & Eloff, J. N. (2017). Anti-inflammatory and anticholinesterase activity of six flavonoids isolated from Polygonum and Dorstenia species. *Archives of Pharmacal Research*, 40(10), 1129–1134. <https://doi.org/10.1007/s12272-015-0612-9>

El Halawany, A. M., El Sayed, N. S., Abdallah, H. M., & El Dine, R. S. (2017). Protective effects of gingerol on streptozotocin-induced sporadic Alzheimer's disease: Emphasis on inhibition of  $\beta$ -amyloid, COX-2, alpha-, beta - secretases and APH1a. *Scientific Reports*, 7(1). <https://doi.org/10.1038/s41598-017-02961-0>

Ellman, G. L., Courtney, K. D., Andres, V., & Featherstone, R. M. (1961). A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochemical Pharmacology*, 7(2), 88–95. [https://doi.org/10.1016/0006-2952\(61\)90145-9](https://doi.org/10.1016/0006-2952(61)90145-9)

Elufioye, T. O., Obuotor, E. M., Agbedahunsi, J. M., & Adesanya, S. A. (2016). Cholinesterase inhibitory activity and structure elucidation of a new phytol derivative and a new cinnamic acid ester from Pycnanthus angolensis. *Revista Brasileira de Farmacognosia*, 26(4), 433–437. <https://doi.org/10.1016/j.bjp.2016.01.010>

Eom, M. R., Weon, J. B., Jung, Y. S., Ryu, G. H., Yang, W. S., & Ma, C. J. (2017). Neuroprotective compounds from Reynoutria sachalinensis. *Archives of Pharmacal Research*, 40(6), 704–712. <https://doi.org/10.1007/s12272-017-0918-x>

Esselun, C., Bruns, B., Hagl, S., Grewal, R., & Eckert, G. P. (2021). Impact of silibinin a on bioenergetics in pc12appsw cells and mitochondrial membrane properties in murine brain mitochondria. *Antioxidants*, 10(10). <https://doi.org/10.3390/antiox10101520>

Farias, J. O., Lima, F. P., Da Conceição, P. dos S., Ribeiro, I. E., Santos, L. G., Da Silva, I. R. T. T., & Silveira, M. S. (2022). A utilização de plantas medicinais no tratamento da doença de alzheimer. *Brazilian Journal of Health Review*, 5(6), 23470–23484. <https://doi.org/10.34119/bjhrv5n6-130>

Ghayur, M. N., Gilani, A. H., Ahmed, T., Khalid, A., Nawaz, S. A., Agbedahunsi, J. M., Choudhary, M. I., & Houghton, P. J. (2008). Muscarinic, Ca<sup>++</sup> antagonist and specific butyrylcholinesterase inhibitory activity of dried ginger extract might explain its use in dementia. *Journal of Pharmacy and Pharmacology*, 60(10), 1375–1383. <https://doi.org/10.1211/jpp/60.10.0014>

Gonzalez, E. P., Hagenow, S., Murillo, M. A., Stark, H., & Suarez, L. C. (2020). Isoquinoline alkaloids from the roots of Zanthoxylum rigidum as multi-target inhibitors of cholinesterase, monoamine oxidase A and A $\beta$ 1-42 aggregation. *Bioorganic Chemistry*, 98. <https://doi.org/10.1016/j.bioorg.2020.103722>

Gríñan-Ferré, C., Bellver-Sanchis, A., Izquierdo, V., Corpas, R., Roig-Soriano, J., Chillón, M., Andres-Lacueva, C., Somogyvári, M., Sóti, C., Sanfeliu, C., & Pallàs, M. (2021). The pleiotropic neuroprotective effects of resveratrol in cognitive decline and Alzheimer's disease pathology: From antioxidant to epigenetic therapy. In *Ageing Research Reviews* (Vol. 67). Elsevier Ireland Ltd. <https://doi.org/10.1016/j.arr.2021.101271>

Guimarães, C. C., Oliveira, D. D., Valdevite, M., Saltoratto, A. L. F., Pereira, S. I. V., França, S. C., Pereira, A. M. S., & Pereira, P. S. (2015). The glycosylated flavonoids vitexin, isovitexin, and querctetin isolated from Serjania erecta Radlk (Sapindaceae) leaves protect PC12 cells against amyloid- $\beta$ 25-35 peptide-induced toxicity. *Food and Chemical Toxicology*, 86, 88–94. <https://doi.org/10.1016/j.fct.2015.09.002>



Guo, A. J. Y., Xie, H. Q., Choi, R. C. Y., Zheng, K. Y. Z., Bi, C. W. C., Xu, S. L., Dong, T. T. X., & Tsim, K. W. K. (2010). Galangin, a flavonol derived from Rhizoma Alpiniae Officinarum, inhibits acetylcholinesterase activity in vitro. *Chemico-Biological Interactions*, 187(1–3), 246–248. <https://doi.org/10.1016/j.cbi.2010.05.002>

Han, R., Yuan, T., Yang, Z., Zhang, Q., Wang, W.-W., Lin, L.-B., Zhu, M.-Q., & Gao, J.-M. (2021). Ulmoidol, an unusual nortriterpenoid from Eucommia ulmoides Oliv. Leaves prevents neuroinflammation by targeting the PU.1 transcriptional signaling pathway. *Bioorganic Chemistry*, 116. <https://doi.org/10.1016/j.bioorg.2021.105345>

Heo, H.-J., Hong, S.-C., Cho, H.-Y., Hong, B., Kim, H.-K., Kim, E.-K., & Shin, D.-H. (2002). Inhibitory effect of zeatin, isolated from Fiatoua villosa, on acetylcholinesterase activity from PC12 cells. *Molecules and Cells*, 13(1), 113–117. <https://www.scopus.com/inward/record.uri?eid=2-s2.0-0037187052&partnerID=40&md5=a7b7afb89e6f001d1a37c44f6eb1db16>

Hoang, T. H. X., Ho, D. V., Van Phan, K., Le, Q. V., Raal, A., & Nguyen, H. T. (2020). Effects of Hippeastrum reticulatum on memory, spatial learning and object recognition in a scopolamine-induced animal model of Alzheimer's disease. *Pharmaceutical Biology*, 58(1), 1098–1104. <https://doi.org/10.1080/13880209.2020.1841810>

Hostalkova, A., Marikova, J., Opletal, L., Korabecny, J., Hulcova, D., Kunes, J., Novakova, L., Perez, D. I., Jun, D., Kucera, T., Andrisano, V., Siatka, T., & Cahlikova, L. (2019). Isoquinoline alkaloids from berberis vulgaris as potential lead compounds for the treatment of alzheimer's disease. *Journal of Natural Products*, 82(2), 239–248. <https://doi.org/10.1021/acs.jnatprod.8b00592>

Hou, W.-C., Lin, R.-D., Chen, C.-T., & Lee, M.-H. (2005). Monoamine oxidase B (MAO-B) inhibition by active principles from Uncaria rhynchophylla. *Journal of Ethnopharmacology*, 100(1–2), 216–220. <https://doi.org/10.1016/j.jep.2005.03.017>

Hu, J., Lin, T., Gao, Y., Xu, J., Jiang, C., Wang, G., Bu, G., Xu, H., Chen, H., & Zhang, Y.-W. (2015). The resveratrol trimer miyabenol C inhibits  $\beta$ -secretase activity and  $\beta$ -amyloid generation. *PLoS ONE*, 10(1). <https://doi.org/10.1371/journal.pone.0115973>

Hu, Q., Wang, J., Irshad, M., Mao, S., Chen, H., Song, Y., Xu, X., & Feng, X. (2022). Neuroprotective Effects of the Psychoactive Compound Biatractylolide (BD) in Alzheimer's Disease. *Molecules*, 27(23). <https://doi.org/10.3390/molecules27238294>

Ibrahim, M., Farooq, T., Hussain, N., Hussain, A., Gulzar, T., Hussain, I., Akash, M. S., & Rehmani, F. S. (2013). Acetyl and butyryl cholinesterase inhibitory sesquiterpene lactones from Amberboa ramosa. *Chemistry Central Journal*, 7(1). <https://doi.org/10.1186/1752-153X-7-116>

Ingkaninan, K., Phengpa, P., Yuenyongsawad, S., & Khorana, N. (2006). Acetylcholinesterase inhibitors from Stephania venosa tuber. *Journal of Pharmacy and Pharmacology*, 58(5), 695–700. <https://doi.org/10.1211/jpp.58.5.0015>

Innok, W., Hiranrat, A., Chana, N., Rungrotmongkol, T., & Kongsune, P. (2021). In silico and in vitro anti-AChE activity investigations of constituents from Mytragyna speciosa for Alzheimer's disease treatment. *Journal of Computer-Aided Molecular Design*, 35(3), 325–336. <https://doi.org/10.1007/s10822-020-00372-4>

Jaipea, S., Saehlim, N., Sutcharitruk, W., Athipornchai, A., Ingkaninan, K., & Saeeng, R. (2023). Synthesis of piperine analogues as AChE and BChE inhibitors for the treatment of Alzheimer's disease. *Phytochemistry Letters*, 53, 216–221. <https://doi.org/10.1016/j.phytol.2023.01.004>



Jamila, N., Yeong, K. K., Murugaiyah, V., Atlas, A., Khan, I., Khan, N., Khan, S. N., Khairuddean, M., & Osman, H. (2015). Molecular docking studies and in vitro cholinesterase enzyme inhibitory activities of chemical constituents of *Garcinia hombroniana*. *Natural Product Research*, 29(1), 86–90. <https://doi.org/10.1080/14786419.2014.952228>

Jamshidi-Kia, F., Lorigooini, Z., & Amini-Khoei, H. (2018). Medicinal plants: Past history and future perspective. Em *Journal of HerbMed Pharmacology* (Vol. 7, Número 1, p. 1–7). Nickan Research Institute. <https://doi.org/10.15171/jhp.2018.01>

Jatav, S., Pandey, N., Dwivedi, P., Akhtar, A., Jyoti, Singh, R., Bansal, R., & Mishra, B. B. (2022). Synthesis of deoxy-Andrographolide Triazolyl Glycoconjugates for the Treatment of Alzheimer's Disease. *ACS Chemical Neuroscience*, 13(23), 3271–3280. <https://doi.org/10.1021/acschemneuro.2c00328>

Jeong, C.-H., Jeong, H. R., Choi, G. N., Kim, D.-O., Lee, U., & Heo, H. J. (2011). Neuroprotective and anti-oxidant effects of caffeic acid isolated from *Erigeron annuus* leaf. *Chinese Medicine*, 6. <https://doi.org/10.1186/1749-8546-6-25>

José Ribeiro Guimarães, A., Sergio da Conceição Moreira, P., & Aparecido Bezerra, C. (2021). MODELOS DE INOVAÇÃO: Análise bibliométrica da produção científica Innovation models: Bibliometric analysis of scientific production. *Brazilian Journal of Information Science: Research trends*, 15, 2106. <https://doi.org/10.36311/1981.1640.2001.v15.e02106>

Jung, H. A., Jung, Y. J., Hyun, S. K., Min, B.-S., Kim, D.-W., Jung, J. H., & Choi, J. S. (2010). Selective cholinesterase inhibitory activities of a new monoterpenoid diglycoside and other constituents from *Nelumbo nucifera* stamens. *Biological and Pharmaceutical Bulletin*, 33(2), 267–272. <https://doi.org/10.1248/bpb.33.267>

Jung, H. A., Lee, E. J., Kim, J. S., Kang, S. S., Lee, J.-H., Min, B.-S., & Choi, J. S. (2009). Cholinesterase and BACE1 inhibitory diterpenoids from *Aralia cordata*. *Archives of Pharmacal Research*, 32(10), 1399–1408. <https://doi.org/10.1007/s12272-009-2009-0>

Jyotshna, Srivastava, N., Singh, B., Chanda, D., & Shanker, K. (2015). Chemical composition and acetylcholinesterase inhibitory activity of *Artemisia maderaspatana* essential oil. *Pharmaceutical Biology*, 53(11), 1677–1683. <https://doi.org/10.3109/13880209.2014.1001405>

Karakoyun, Ç., Bozkurt, B., Çoban, G., Masi, M., Cimmino, A., Evidente, A., & Unver Somer, N. (2020). A comprehensive study on *narcissus tazetta* subsp. *tazetta* L.: Chemo-profiling, isolation, anticholinesterase activity and molecular docking of amaryllidaceae alkaloids. *South African Journal of Botany*, 130, 148–154. <https://doi.org/10.1016/j.sajb.2019.11.016>

Kareti, S. R., & Pharm, S. M. (2020). In Silico Molecular Docking Analysis of Potential Anti-Alzheimer's Compounds Present in Chloroform Extract of *Carissa carandas* Leaf Using Gas Chromatography MS/MS. *Current Therapeutic Research - Clinical and Experimental*, 93. <https://doi.org/10.1016/j.curtheres.2020.100615>

Karim, N., Khan, I., Abdelhalim, A., Abdel-Halim, H., & Hanrahan, J. R. (2017). Molecular docking and antiamnesic effects of nepitrin isolated from *Rosmarinus officinalis* on scopolamine-induced memory impairment in mice. *Biomedicine and Pharmacotherapy*, 96, 700–709. <https://doi.org/10.1016/j.biopha.2017.09.121>



Kasote, D. M., Katyare, S. S., Hegde, M. V., & Bae, H. (2015). Significance of antioxidant potential of plants and its relevance to therapeutic applications. Em *International Journal of Biological Sciences* (Vol. 11, Número 8, p. 982–991). Ivyspring International Publisher. <https://doi.org/10.7150/ijbs.12096>

Khatami, Z., Herdlinger, S., Sarkhail, P., Zehl, M., Kaehlig, H., Schuster, D., & Adhami, H.-R. (2020). Isolation and characterization of acetylcholinesterase inhibitors from piper longum and binding mode predictions. *Planta Medica*, 86(15), 1118–1124. <https://doi.org/10.1055/a-1199-7084>

Kim, J. W., Seo, J. Y., Oh, W. K., & Sung, S. H. (2017). Anti-neuroinflammatory ent-kaurane diterpenoids from Pteris multifida roots. *Molecules*, 22(1). <https://doi.org/10.3390/molecules22010027>

Kim, J. Y., Lee, W. S., Kim, Y. S., Curtis-Long, M. J., Lee, B. W., Ryu, Y. B., & Park, K. H. (2011). Isolation of cholinesterase-inhibiting flavonoids from Morus lhou. *Journal of Agricultural and Food Chemistry*, 59(9), 4589–4596. <https://doi.org/10.1021/jf200423g>

Kim, Y. J., Sohn, E., Kim, J.-H., Na, M., & Jeong, S.-J. (2020). Catechol-Type Flavonoids from the Branches of Elaeagnus glabra f. oxyphylla Exert Antioxidant Activity and an Inhibitory Effect on Amyloid- $\beta$  Aggregation. *Molecules (Basel, Switzerland)*, 25(21). <https://doi.org/10.3390/molecules25214917>

Koirala, P., Seong, S. H., Jung, H. A., & Choi, J. S. (2017). Comparative molecular docking studies of lupeol and lupenone isolated from Pueraria lobata that inhibits BACE1: Probable remedies for Alzheimer's disease. *Asian Pacific Journal of Tropical Medicine*, 10(12), 1117–1122. <https://doi.org/10.1016/j.apjtm.2017.10.018>

Krishnan, N., Mariappanadar, V., Dhanabalan, A. K., Devadasan, V., Gopinath, S. C. B., & Raman, P. (2022). Purification, identification and in silico models of alkaloids from Nardostachys jatamansi — bioactive compounds for neurodegenerative diseases. *Biomass Conversion and Biorefinery*. <https://doi.org/10.1007/s13399-022-03237-y>

Kurisu, M., Miyamae, Y., Murakami, K., Han, J., Isoda, H., Irie, K., & Shigemori, H. (2013). Inhibition of amyloid  $\beta$  aggregation by acteoside, a phenylethanoid glycoside. *Bioscience, Biotechnology and Biochemistry*, 77(6), 1329–1332. <https://doi.org/10.1271/bbb.130101>

Kurutas, E. B. (2016). The importance of antioxidants which play the role in cellular response against oxidative/nitrosative stress: Current state. Em *Nutrition Journal* (Vol. 15, Número 1). BioMed Central Ltd. <https://doi.org/10.1186/s12937-016-0186-5>

Kwon, H. C., Cha, J. W., Park, J.-S., Chun, Y. S., Moodley, N., Maharaj, V. J., Youn, S. H., Chung, S., & Yang, H. O. (2011). Rapid identification of bioactive compounds reducing the production of amyloid  $\beta$ -peptide ( $A\beta$ ) from south African plants using an automated HPLC/SPE/HPLC coupling system. *Biomolecules and Therapeutics*, 19(1), 90–96. <https://doi.org/10.4062/biomolther.2011.19.1.090>

Lai, D.-H., Yang, Z.-D., Xue, W.-W., Sheng, J., Shi, Y., & Yao, X.-J. (2013). Isolation, characterization and acetylcholinesterase inhibitory activity of alkaloids from roots of Stemona sessilifolia. *Fitoterapia*, 89(1), 257–264. <https://doi.org/10.1016/j.fitote.2013.06.010>

Lam, L. M. T., Nguyen, M. T. T., Nguyen, H. X., Dang, P. H., Nguyen, N. T., Tran, H. M., Nguyen, H. T., Nguyen, N. M., Min, B. S., Kim, J. A., Choi, J. S., & Can, M. (2016). Anti-cholinesterases and memory improving effects of Vietnamese Xylia xylocarpa. *Chemistry Central Journal*, 10(1). <https://doi.org/10.1186/s13065-016-0197-5>



Lauer, A. A., Grimm, H. S., Apel, B., Golobrodska, N., Kruse, L., Ratanski, E., Schulten, N., Schwarze, L., Slawik, T., Sperlich, S., Vohla, A., & Grimm, M. O. W. (2022). Mechanistic Link between Vitamin B12 and Alzheimer's Disease. Em *Biomolecules* (Vol. 12, Número 1). MDPI. <https://doi.org/10.3390/biom12010129>

Lawal, B. A., Udobre, A., Elufioye, T. O., Ahmadu, A. A., & Olanipekun, B. (2020). Novel cholinesterase inhibitory effect of  $\alpha$ -spinasterol isolated from the leaves of *Acacia auriculiformis* A. CUNN Ex. Benth (Fabaceae). *Tropical Journal of Pharmaceutical Research*, 19(7), 1473–1480. <https://doi.org/10.4314/tjpr.v19i7.20>

Lee, D.-S., Cha, B.-Y., Woo, J.-T., Kim, Y.-C., & Jang, J.-H. (2015). Acerogenin A from *Acer nikoense* Maxim prevents oxidative stress-induced neuronal cell death through Nrf2-mediated heme oxygenase-1 expression in mouse hippocampal HT22 cell line. *Molecules*, 20(7), 12545–12557. <https://doi.org/10.3390/molecules200712545>

Lee, D.-S., Lee, M., Sung, S. H., & Jeong, G. S. (2016). Involvement of heme oxygenase-1 induction in the cytoprotective and neuroinflammatory activities of Siegesbeckia Pubescens isolated from 5,3'-dihydroxy-3,7,4'-trimethoxyflavone in HT22 cells and BV2 cells. *International Immunopharmacology*, 40, 65–72. <https://doi.org/10.1016/j.intimp.2016.08.030>

Lee, H. W., Ryu, H. W., Kang, M.-G., Park, D., Lee, H., Shin, H. M., Oh, S.-R., & Kim, H. (2017). Potent inhibition of monoamine oxidase A by decursin from *Angelica gigas* Nakai and by wogonin from *Scutellaria baicalensis* Georgi. *International Journal of Biological Macromolecules*, 97, 598–605. <https://doi.org/10.1016/j.ijbiomac.2017.01.080>

Lee, H. W., Ryu, H. W., Kang, M.-G., Park, D., Oh, S.-R., & Kim, H. (2016). Potent selective monoamine oxidase B inhibition by maackiain, a pterocarpan from the roots of *Sophora flavescens*. *Bioorganic and Medicinal Chemistry Letters*, 26(19), 4714–4719. <https://doi.org/10.1016/j.bmcl.2016.08.044>

Lee, H.-J., Park, E.-J., Lee, B.-W., Cho, H.-M., Pham, T.-L.-G., Hoang, Q.-H., Pan, C.-H., & Oh, W.-K. (2021). Flavanonol glycosides from the stems of *myrsine seguinii* and their neuroprotective activities. *Pharmaceuticals*, 14(9). <https://doi.org/10.3390/ph14090911>

Lee, Y. K., Bang, H. J., Oh, J. B., & Whang, W. K. (2017). Bioassay-Guided isolated compounds from *morinda officinalis* inhibit Alzheimer's disease pathologies. *Molecules*, 22(10). <https://doi.org/10.3390/molecules22101638>

Lee, Y. K., Yuk, D. Y., Kim, T. I., Kim, Y. H., Kim, K. T., Kim, K. H., Lee, B. J., Nam, S.-Y., & Hong, J. T. (2009). Protective effect of the ethanol extract of *Magnolia officinalis* and 4-O-methylhonokiol on scopolamine-induced memory impairment and the inhibition of acetylcholinesterase activity. *Journal of Natural Medicines*, 63(3), 274–282. <https://doi.org/10.1007/s11418-009-0330-z>

Li, C.-H., Meng, X.-H., Lan Huong, D., Oh, W. K., Wang, W.-F., & Yang, J.-L. (2019). Quantitative visualization and detection of acetylcholinesterase activity and its inhibitor based on the oxidation character of ultrathin MnO<sub>2</sub> nanosheets. *Analytical Methods*, 11(38), 4931–4938. <https://doi.org/10.1039/c9ay01721f>

Li, F.-J., Liu, Y., Yuan, Y., Yang, B., Liu, Z.-M., & Huang, L.-Q. (2017). Molecular interaction studies of acetylcholinesterase with potential acetylcholinesterase inhibitors from the root of *Rhodiola crenulata* using molecular docking and isothermal titration calorimetry methods. *International Journal of Biological Macromolecules*, 104, 527–532. <https://doi.org/10.1016/j.ijbiomac.2017.06.066>



Li, S., Liu, C., Zhang, Y., & Tsao, R. (2021). On-line coupling pressurised liquid extraction with two-dimensional counter current chromatography for isolation of natural acetylcholinesterase inhibitors from *Astragalus membranaceus*. *Phytochemical Analysis*, 32(4), 640–653. <https://doi.org/10.1002/pca.3012>

Li, W., Lu, Q., Li, X., Liu, H., Sun, L., Lu, X., Zhao, Y., & Liu, P. (2020). Anti-Alzheimer's disease activity of secondary metabolites from: *Xanthoceras sorbifolia* Bunge. *Food and Function*, 11(3), 2067–2079. <https://doi.org/10.1039/c9fo01138b>

Lim, H. J., Li, H., Kim, J. Y., & Ryu, J.-H. (2011). Quercetin derivatives from *Siegesbeckia glabrescens* inhibit the expression of COX-2 through the suppression of NF-κB activation in microglia. *Biomolecules and Therapeutics*, 19(1), 27–32. <https://doi.org/10.4062/biomolther.2011.19.1.027>

Liu, L., Yin, Q.-M., Gao, Q., Li, J., Jiang, Y., & Tu, P.-F. (2021). New biphenanthrenes with butyrylcholinesterase inhibitory activity from *Cremastra appendiculata*. *Natural Product Research*, 35(5), 750–756. <https://doi.org/10.1080/14786419.2019.1601091>

Liu, Y.-M., Feng, Y.-D., Lu, X., Nie, J.-B., Li, W., Wang, L.-N., Tian, L.-J., & Liu, Q.-H. (2017). Isosteroidal alkaloids as potent dual-binding site inhibitors of both acetylcholinesterase and butyrylcholinesterase from the bulbs of *Fritillaria walujewii*. *European Journal of Medicinal Chemistry*, 137, 280–291. <https://doi.org/10.1016/j.ejmech.2017.06.007>

Machado, A. P. R., Carvalho, I. O., & Rocha Sobrinho, H. M. da. (2020). NEUROINFLAMAÇÃO NA DOENÇA DE ALZHEIMER. *Revista Brasileira Militar de Ciências*, 6(14). <https://doi.org/10.36414/rbmc.v6i14.33>

Mahnashi, M. H., Alqahtani, Y. S., Alqarni, A. O., Alyami, B. A., Jan, M. S., Ayaz, M., Ullah, F., Rashid, U., & Sadiq, A. (2021). Crude extract and isolated bioactive compounds from *Notholirion thomsonianum* (Royale) Stapf as multitargets antidiabetic agents: in-vitro and molecular docking approaches. *BMC complementary medicine and therapies*, 21(1), 270. <https://doi.org/10.1186/s12906-021-03443-7>

Mahnashi, M. H., Alshahrani, M. A., Nahari, M. H., Hassan, S. S. U., Jan, M. S., Ayaz, M., Ullah, F., Alshehri, O. M., Alshehri, M. A., Rashid, U., & Sadiq, A. (2022). In-Vitro, In-Vivo, Molecular Docking and ADMET Studies of 2-Substituted 3,7-Dihydroxy-4H-chromen-4-one for Oxidative Stress, Inflammation and Alzheimer's Disease. *Metabolites*, 12(11). <https://doi.org/10.3390/metabo12111055>  
Mahnashi, M. H., & Alshehri, O. M. (2022). Isolation, In Vitro and In Silico Anti-Alzheimer and Anti-Inflammatory Studies on Phytosteroids from Aerial Parts of *Fragaria × ananassa* Duch. *Biomolecules*, 12(10). <https://doi.org/10.3390/biom12101430>

Mahnashi, M. H., Alyami, B. A., Alqahtani, Y. S., Alqarni, A. O., Jan, M. S., Hussain, F., Zafar, R., Rashid, U., Abbas, M., Tariq, M., & Sadiq, A. (2022). Antioxidant Molecules Isolated from Edible Prostrate Knotweed: Rational Derivatization to Produce More Potent Molecules. *Oxidative Medicine and Cellular Longevity*, 2022. <https://doi.org/10.1155/2022/3127480>

Mahran, E., Morlock, G. E., & Keusgen, M. (2020). Guided isolation of new iridoid glucosides from *Anarrhinum pubescens* by high-performance thin-layer chromatography-acetylcholinesterase assay. *Journal of Chromatography A*, 1609. <https://doi.org/10.1016/j.chroma.2019.460438>

Mai, Y., Wang, Z., Wang, Y., Xu, J., & He, X. (2020). Anti-neuroinflammatory triterpenoids from the seeds of *Quercus serrata* Thunb. *Fitoterapia*, 142. <https://doi.org/10.1016/j.fitote.2020.104523>



Marucci, G., Buccioni, M., Ben, D. D., Lambertucci, C., Volpini, R., & Amenta, F. (2021). Efficacy of acetylcholinesterase inhibitors in Alzheimer's disease. In *Neuropharmacology* (Vol. 190). Elsevier Ltd. <https://doi.org/10.1016/j.neuropharm.2020.108352>

Masondo, N. A., Stafford, G. I., Aremu, A. O., & Makunga, N. P. (2019). Acetylcholinesterase inhibitors from southern African plants: An overview of ethnobotanical, pharmacological potential and phytochemical research including and beyond Alzheimer's disease treatment. In *South African Journal of Botany* (Vol. 120, p. 39–64). Elsevier B.V. <https://doi.org/10.1016/j.sajb.2018.09.011>

Min, B. S., Cuong, T. D., Lee, J.-S., Shin, B.-S., Woo, M. H., & Hung, T. M. (2010). Cholinesterase inhibitors from Cleistocalyx operculatus buds. *Archives of Pharmacal Research*, 33(10), 1665–1670. <https://doi.org/10.1007/s12272-010-1016-5>

Mira, A., Yamashita, S., Katakura, Y., & Shimizu, K. (2015). In vitro neuroprotective activities of compounds from Angelica shikokiana Makino. *Molecules*, 20(3), 4813–4832. <https://doi.org/10.3390/molecules20034813>

Moon, U. R., Sircar, D., Barthwal, R., Sen, S. K., Beuerle, T., Beerhues, L., & Mitra, A. (2015). Shoot cultures of Hoppea fastigiata (Griseb.) C.B. Clarke as potential source of neuroprotective xanthones. *Journal of Natural Medicines*, 69(3), 375–386. <https://doi.org/10.1007/s11418-015-0904-x>

Muñoz-Nuñez, E., Quiroz-Carreño, S., Pastene-Navarrete, E., Seigler, D. S., Céspedes-Acuña, C., Martínez Valenzuela, I., Oppiger Muñoz, M., Salas-Burgos, A., & Alarcón-Enos, J. (2022). Assessments of Ceanothanes Triterpenes as Cholinesterase Inhibitors: An Investigation of Potential Agents with Novel Inspiration for Drug Treatment of Neurodegenerative Diseases. *Metabolites*, 12(7). <https://doi.org/10.3390/metabo12070668>

Na, C. S., Hong, S. S., Choi, Y.-H., Lee, Y. H., Hong, S. H., Lim, J.-Y., Kang, B. H., Park, S.-Y., & Lee, D. (2010). Neuroprotective effects of constituents of Eragrostis ferruginea against A $\beta$ -induced toxicity in PC12 cells. *Archives of Pharmacal Research*, 33(7), 999–1003. <https://doi.org/10.1007/s12272-010-0704-5>

Nair, J. J., Aremu, A. O., & Van Staden, J. (2011). Isolation of narciprimine from Cyrtanthus contractus (Amaryllidaceae) and evaluation of its acetylcholinesterase inhibitory activity. *Journal of Ethnopharmacology*, 137(3), 1102–1106. <https://doi.org/10.1016/j.jep.2011.07.028>

Namdaung, U., Athipornchai, A., Khammee, T., Kuno, M., & Suksamrarn, S. (2018). 2-Arylbenzofurans from Artocarpus lakoocha and methyl ether analogs with potent cholinesterase inhibitory activity. *European Journal of Medicinal Chemistry*, 143, 1301–1311. <https://doi.org/10.1016/j.ejmech.2017.10.019>

Nassief, S. M., Amer, M. E., Shawky, E., Saleh, S. R., & El-Masry, S. (2020). Acetylcholinesterase Inhibitory Alkaloids from the Flowers and Seeds of Erythrina caffra. *Revista Brasileira de Farmacognosia*, 30(6), 859–864. <https://doi.org/10.1007/s43450-020-00114-5>

Nguyen, T. K., Tran, T. H., Nguyen, K., Ho, D. V., Nguyen, H. T., & Tran, L. T. T. (2022). Deep Learning Model to Identify Potential Acetylcholinesterase Inhibitors: A Case Study of Isolated Compounds From Pongamia pinnata (L.) Pierre. *Natural Product Communications*, 17(7). <https://doi.org/10.1177/1934578X221117310>

Nugroho, A., Choi, J. S., Hong, J.-P., & Park, H.-J. (2017). Anti-acetylcholinesterase activity of the aglycones of phenolic glycosides isolated from Leonurus japonicus. *Asian Pacific Journal of Tropical Biomedicine*, 7(10), 849–854. <https://doi.org/10.1016/j.apjtb.2017.08.013>



Nugroho, A., Choi, J. S., Seong, S. H., Song, B.-M., Park, K.-S., & Park, H.-J. (2018). Isolation of flavonoid glycosides with cholinesterase inhibition activity and quantification from stachys japonica. *Natural Product Sciences*, 24(4), 259–265. <https://doi.org/10.20307/NPS.2018.24.4.259>

Nugroho, A., Park, J.-H., Choi, J. S., Park, K.-S., Hong, J.-P., & Park, H.-J. (2019). Structure determination and quantification of a new flavone glycoside with anti-acetylcholinesterase activity from the herbs of Elsholtzia ciliata. *Natural Product Research*, 33(6), 814–821. <https://doi.org/10.1080/14786419.2017.1413556>

Oh, J. M., Jang, H.-J., Kang, M.-G., Song, S., Kim, D.-Y., Kim, J.-H., Noh, J.-I., Park, J. E., Park, D., Yee, S.-T., & Kim, H. (2021). Acetylcholinesterase and monoamine oxidase-B inhibitory activities by ellagic acid derivatives isolated from Castanopsis cuspidata var. sieboldii. *Scientific Reports*, 11(1), 13953. <https://doi.org/10.1038/s41598-021-93458-4>

Onoja, O. J., Elufioye, T. O., Sherwani, Z. A., & Ul-Haq, Z. (2020). Molecular Docking Studies and Anti-Alzheimer's Potential of Isolated Compounds from Tinospora cordifolia. *Journal of Biologically Active Products from Nature*, 10(2), 100–121. <https://doi.org/10.1080/22311866.2020.1726813>

Orhan, I. E., Khan, M. T. H., Erdem, S. A., Kartal, M., & Şener, B. (2011). Selective cholinesterase inhibitors from Buxus sempervirens L. and their molecular docking studies. *Current Computer-Aided Drug Design*, 7(4), 276–286. <https://doi.org/10.2174/157340911798260296>

Orhan, I. E., Kucukboyaci, N., Calis, I., Cerón-Carrasco, J. P., den-Haan, H., Peña-García, J., & Pérez-Sánchez, H. (2017). Acetylcholinesterase inhibitory assessment of isolated constituents from Salsola grandis Freitag, Vural & Adıguzel and molecular modeling studies on N-acetyltryptophan. *Phytochemistry Letters*, 20, 373–378. <https://doi.org/10.1016/j.phytol.2016.10.017>

Orhan, I. E., Senol, F. S., Shekfeh, S., Skalicka-Wozniak, K., & Banoglu, E. (2017). Pteryxin - A promising butyrylcholinesterase-inhibiting coumarin derivative from Mutellina purpurea. *Food and Chemical Toxicology*, 109, 970–974. <https://doi.org/10.1016/j.fct.2017.03.016>

Ortiz, J. E., Garro, A., Pigni, N. B., Agüero, M. B., Roitman, G., Slanis, A., Enriz, R. D., Feresin, G. E., Bastida, J., & Tapia, A. (2018). Cholinesterase-inhibitory effect and in silico analysis of alkaloids from bulbs of Hieronymiella species. *Phytomedicine*, 39, 66–74. <https://doi.org/10.1016/j.phymed.2017.12.020>

Othman, A., Sayed, A. M., Amen, Y., & Shimizu, K. (2022). Possible neuroprotective effects of amide alkaloids from Bassia indica and Agathophora alopecuroides: in vitro and in silico investigations. *RSC Advances*, 12(29), 18746–18758. <https://doi.org/10.1039/d2ra02275c>

Özaslan, M. S., Sağlamtaş, R., Demir, Y., Genç, Y., Saracoğlu, İ., & Gülçin, İ. (2022). Isolation of Some Phenolic Compounds from Plantago subulata L. and Determination of Their Antidiabetic, Anticholinesterase, Antiepileptic and Antioxidant Activity. *Chemistry and Biodiversity*, 19(8). <https://doi.org/10.1002/cbdv.202200280>

Panche, A. N., Chandra, S., & Diwan, A. D. (2019). Multi-target  $\beta$ -protease inhibitors from andrographis paniculata: In silico and in vitro studies. *Plants*, 8(7). <https://doi.org/10.3390/plants8070231>

Panda, S. S., & Jhanji, N. (2020). Natural products as potential anti-alzheimer agents. *Current Medicinal Chemistry*, 27(35), 5887–5917. <https://doi.org/10.2174/0929867326666190618113613>



Park, J. H., & Whang, W. K. (2020). Bioassay-Guided Isolation of Anti-Alzheimer Active Components from the Aerial Parts of *Hedyotis diffusa* and Simultaneous Analysis for Marker Compounds. *Molecules*, 25(24). <https://doi.org/10.3390/MOLECULES25245867>

Park, S.-Y., & Kim, D. S. H. L. (2002). Discovery of natural products from *Curcuma longa* that protect cells from beta-amyloid insult: A drug discovery effort against Alzheimer's disease. *Journal of Natural Products*, 65(9), 1227–1231. <https://doi.org/10.1021/np010039x>

Peng, X.-R., Wang, X., Dong, J.-R., Qin, X.-J., Li, Z.-R., Yang, H., Zhou, L., & Qiu, M.-H. (2017). Rare Hybrid Dimers with Anti-Acetylcholinesterase Activities from a Safflower (*Carthamus tinctorius* L.) Seed Oil Cake. *Journal of Agricultural and Food Chemistry*, 65(43), 9453–9459. <https://doi.org/10.1021/acs.jafc.7b03431>

Pitchai, A., Nagarajan, N., Vincent, S. G. P., & Rajaretnam, R. K. (2018). Zebrafish bio-assay guided isolation of human acetylcholinesterase inhibitory trans-tephrostachin from *Tephrosia purpurea* (L.) Pers. *Neuroscience Letters*, 687, 268–275. <https://doi.org/10.1016/j.neulet.2018.09.058>

Posri, P., Suthiwong, J., Takomthong, P., Wongsa, C., Chuenban, C., Boonyarat, C., & Yenjai, C. (2019). A new flavonoid from the leaves of *Atalantia monophylla* (L.) DC. *Natural Product Research*, 33(8), 1115–1121. <https://doi.org/10.1080/14786419.2018.1457667>

Rahman, A.-U., Khalid, A., Sultana, N., Nabeel Ghayur, M., Ahmed Mesaik, M., Riaz Khan, M., Gilani, A. H., & Iqbal Choudhary, M. (2006). New natural cholinesterase inhibiting and calcium channel blocking quinoline alkaloids. *Journal of Enzyme Inhibition and Medicinal Chemistry*, 21(6), 703–710. <https://doi.org/10.1080/14756360600889708>

Rajput, A., Sharma, P., Kumar, N., Kaur, S., & Arora, S. (2023). Neuroprotective activity of novel phenanthrene derivative from *Grewia tiliaceaefolia* by in vitro and in silico studies. *Scientific Reports*, 13(1). <https://doi.org/10.1038/s41598-023-29446-7>

Rakkhittawattana, V., Panichayupakaranant, P., Prasanth, M. I., Brimson, J. M., & Tencomnao, T. (2022). Rhinacanthin-C but Not-D Extracted from *Rhinacanthus nasutus* (L.) Kurz Offers Neuroprotection via ERK, CHOP, and LC3B Pathways. *Pharmaceuticals*, 15(5). <https://doi.org/10.3390/ph15050627>

Rashed, K. N. Z., Said, A., Feitosa, C., & Sucupira, A. C. C. (2015). Evaluation of anti-alzheimer activity of *Ampelopsis brevipedunculata* and the isolated compounds. *Research Journal of Phytochemistry*, 9(1), 16–24. <https://doi.org/10.3923/rjphyto.2015.16.24>

Rauter, A. P., Branco, I., Lopes, R. G., Justino, J., Silva, F. V. M., Noronha, J. P., Cabrita, E. J., Brouard, I., & Bermejo, J. (2007). A new lupene triterpenetriol and anticholinesterase activity of *Salvia sclareaoides*. *Fitoterapia*, 78(7–8), 474–481. <https://doi.org/10.1016/j.fitote.2007.02.013>

Rehman, F.-U., Khan, M. F., Khan, I., & Roohullah. (2013). Molecular interactions of an alkaloid euchrestifoline as a new acetylcholinesterase inhibitor. *Bangladesh Journal of Pharmacology*, 8(4), 361–364. <https://doi.org/10.3329/bjp.v8i3.16417>

Ren, R., Qi, J., Lin, S., Liu, X., Yin, P., Wang, Z., Tang, R., Wang, J., Huang, Q., Li, J., Xie, X., Hu, Y., Cui, S., Zhu, Y., Yu, X., Wang, P., Zhu, Y., Wang, Y., Huang, Y., ... Wang, G. (2022). The China Alzheimer Report 2022. *General Psychiatry*, 35(1), e100751. <https://doi.org/10.1136/gpsych-2022-100751>



Ren, Y., Houghton, P. J., Hider, R. C., & Howes, M.-J. R. (2004). Novel diterpenoid acetylcholinesterase inhibitors from *Salvia miltiorrhiza*. *Planta Medica*, 70(3), 201–204. <https://doi.org/10.1055/s-2004-815535>

Rhee, I. K., van Rijn, R. M., & Verpoorte, R. (2003). Qualitative determination of false-positive effects in the acetylcholinesterase assay using thin layer chromatography. *Phytochemical Analysis*, 14(3), 127–131. <https://doi.org/10.1002/pca.675>

Rios, M. Y., Ocampo-Acuña, Y. D., Ramírez-Cisneros, M. Á., & Salazar-Rios, M. E. (2020). Eurofuranone Lignans from *Leucophyllum ambiguum*. *Journal of Natural Products*, 83(5), 1424–1431. <https://doi.org/10.1021/acs.jnatprod.9b00759>

Rodrigues, M. J., Gangadhar, K. N., Zengin, G., Mollica, A., Varela, J., Barreira, L., & Custódio, L. (2017). Juncaceae species as sources of innovative bioactive compounds for the food industry: In vitro antioxidant activity, neuroprotective properties and in silico studies. *Food and Chemical Toxicology*, 107, 590–596. <https://doi.org/10.1016/j.fct.2017.04.006>

Rodrigues, V. G., Silva, F. C., Duarte, L. P., Takahashi, J. A., Matildes, B. L. G., Silva, G. D. F., Silva, R. R., & Vieira-Filho, S. A. (2014). Pentacyclic triterpenes from *Maytenus* genus as acetylcholinesterase inhibitors. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6(SUPPL. 2), 918–920. <https://www.scopus.com/inward/record.uri?eid=2-s2.0-84896479817&partnerID=40&md5=cbe097c53232a49d77983fa347584013>

Saadullah, M., Farid, A., Ali, A., Rashad, M., Naseem, F., Rashid, S. A., Ghazanfar, S., Yasin, M., Akhtar, N., Almuhayawi, M. S., Alruhaili, M. H., & Selim, S. (2022). Molecular Modeling Study of Novel Lancifolamide Bioactive Molecule as an Inhibitor of Acetylcholinesterase (AChE), Herpes Simplex Virus (HSV-1), and Anti-proliferative Proteins. *Molecules*, 27(17). <https://doi.org/10.3390/molecules27175480>

Šafratová, M., Hostalková, A., Hulcová, D., Breiterová, K., Hrabcová, V., Machado, M., Fontinha, D., Prudêncio, M., Kuneš, J., Chlebek, J., Jun, D., Hrabinová, M., Nováková, L., Havelek, R., Seifrtová, M., Opletal, L., & Cahliková, L. (2018). Alkaloids from *Narcissus poeticus* cv. Pink Parasol of various structural types and their biological activity. *Archives of Pharmacal Research*, 41(2), 208–218. <https://doi.org/10.1007/s12272-017-1000-4>

Sawasdee, P., Sabphon, C., Sitthiwongwanit, D., & Kokpol, U. (2009). Anticholinesterase activity of 7-methoxyflavones isolated from *Kaempferia parviflora*. *Phytotherapy Research*, 23(12), 1792–1794. <https://doi.org/10.1002/ptr.2858>

Seo, S.-H., Lee, Y.-C., & Moon, H.-I. (2017). Acetyl-cholinesterase inhibitory activity of methoxyflavones isolated from *Kaempferia parviflora*. *Natural Product Communications*, 12(1), 21–22. <https://doi.org/10.1177/1934578x1701200107>

Sevindik, H. G., Güvenalp, Z., Yerdelen, K. Ö., Yuca, H., & Demirezer, L. Ö. (2015). Research on drug candidate anticholinesterase molecules from *Achillea biebersteinii* Afan. using by molecular docking and in vitro methods. *Medicinal Chemistry Research*, 24(11), 3794–3802. <https://doi.org/10.1007/s00044-015-1423-8>

Shiao, Y.-J., Wang, C.-N., Wang, W.-Y., & Lin, Y.-L. (2005). Neuroprotective flavonoids from *Flemingia macrophylla*. *Planta Medica*, 71(9), 835–840. <https://doi.org/10.1055/s-2005-871297>  
Shrestha, S., Seong, S. H., Paudel, P., Jung, H. A., & Choi, J. S. (2018). Structure related inhibition of enzyme systems in cholinesterases and BACE1 in vitro by naturally occurring naphthopyrone and its



glycosides isolated from cassia obtusifolia. *Molecules*, 23(1).  
<https://doi.org/10.3390/molecules23010069>

Siatka, T., Adamcová, M., Opletal, L., Cahlíková, L., Jun, D., Hrabinová, M., Kuneš, J., & Chlebek, J. (2017). Cholinesterase and prolyl oligopeptidase inhibitory activities of alkaloids from argemone platyceras (Papaveraceae). *Molecules*, 22(7). <https://doi.org/10.3390/molecules22071181>

Sibanyoni, M. N., Chaudhary, S. K., Chen, W., Adhami, H.-R., Combrinck, S., Maharaj, V., Schuster, D., & Viljoen, A. (2020). Isolation, in vitro evaluation and molecular docking of acetylcholinesterase inhibitors from South African Amaryllidaceae. *Fitoterapia*, 146. <https://doi.org/10.1016/j.fitote.2020.104650>

Sichaem, J., Rojpitikul, T., Sawasdee, P., Lugsanangarm, K., & Tip-Pyang, S. (2015). Furoquinoline alkaloids from the leaves of evodia lepta as potential cholinesterase inhibitors and their molecular docking. *Natural Product Communications*, 10(8), 1359–1362. <https://doi.org/10.1177/1934578x1501000811>

Sichaem, J., Tip-Pyang, S., & Lugsanangarm, K. (2018). Bioactive aporphine alkaloids from the roots of Artabotrys spinosus: Cholinesterase inhibitory activity and molecular docking studies. *Natural Product Communications*, 13(10), 1279–1282. <https://doi.org/10.1177/1934578x1801301011>

Simon, A., Darcsi, A., Kéry, Á., & Riethmüller, E. (2020). Blood-brain barrier permeability study of ginger constituents. *Journal of Pharmaceutical and Biomedical Analysis*, 177. <https://doi.org/10.1016/j.jpba.2019.112820>

Singh, V., Kaur, K., Kaur, S., Shri, R., Singh, T. G., & Singh, M. (2022). Trimethoxyflavones from Ocimum basilicum L. leaves improve long term memory in mice by modulating multiple pathways. *Journal of Ethnopharmacology*, 295. <https://doi.org/10.1016/j.jep.2022.115438>

Sivaraman, D., Panneerselvam, P., & Muralidharan, P. (2014). Isolation, characterization and insilico pharmacological screening of medicinally important bio-active phytoconstituents from the leaves of Ipomoea aquatica forsk. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6(2), 262–267. <https://www.scopus.com/inward/record.uri?eid=2-s2.0-84897143031&partnerID=40&md5=17a47db36b7c9ccd47515145c6422af3>

Stefanello, F. M. (2003). Efeito da homocisteína sobre as atividades da butirilcolinesterase e da Na<sup>+</sup>, K<sup>+</sup>-ATPase em sangue de ratos. *Universidade Federal do Rio Grande do Sul. Instituto de Ciências Básicas da Saúde. Programa de Pós-Graduação em Ciências Biológicas: Bioquímica*.

Sun, J., Jiang, G., & Shigemori, H. (2019). Inhibitory Activity on Amyloid Aggregation of Rosmarinic Acid and Its Substructures from Isodon japonicus. *Natural Product Communications*, 14(5). <https://doi.org/10.1177/1934578X19843039>

'T Hart, B. A., Copray, S., & Philippens, I. (2014). Apocynin, a low molecular oral treatment for neurodegenerative disease. *BioMed Research International*, 2014. <https://doi.org/10.1155/2014/298020>

Tang, B., Zeng, W., Song, L. L., Wang, H. M., Qu, L. Q., Lo, H. H., Yu, L., Wu, A. G., Wong, V. K. W., & Law, B. Y. K. (2022). Extracellular Vesicle Delivery of Neferine for the Attenuation of Neurodegenerative Disease Proteins and Motor Deficit in an Alzheimer's Disease Mouse Model. *Pharmaceuticals*, 15(1). <https://doi.org/10.3390/ph15010083>



Teh, S. S., Ee, G. C. L., Mah, S. H., & Ahmad, Z. (2016). Structure-activity relationship study of secondary metabolites from *Mesua beccariana*, *Mesua ferrea* and *Mesua congestiflora* for anti-cholinesterase activity. *Medicinal Chemistry Research*, 25(5), 819–823. <https://doi.org/10.1007/s00044-016-1531-0>

Tettevi, E. J., Maina, M., Simpong, D. L., Osei-Atweneboana, M. Y., & Ocloo, A. (2022). A Review of African Medicinal Plants and Functional Foods for the Management of Alzheimer's Disease-related Phenotypes, Treatment of HSV-1 Infection and/or Improvement of Gut Microbiota. *Journal of Evidence-Based Integrative Medicine*, 27. <https://doi.org/10.1177/2515690X221114657>

Thakur, A., Moyo, P., van der Westhuizen, C. J., Yang, H. O., & Maharaj, V. (2021). A novel cardenolide glycoside isolated from *xysmalobium undulatum* reduces levels of the alzheimer's disease-associated  $\beta$ -amyloid peptides  $\alpha\beta42$  in vitro. *Pharmaceuticals*, 14(8). <https://doi.org/10.3390/ph14080743>

Tiang, N., Ahad, M. A., Murugaiyah, V., & Hassan, Z. (2020). Xanthone-enriched fraction of *Garcinia mangostana* and  $\alpha$ -mangostin improve the spatial learning and memory of chronic cerebral hypoperfusion rats. *Journal of Pharmacy and Pharmacology*, 72(11), 1629–1644. <https://doi.org/10.1111/jphp.13345>

Tuzimski, T., & Petruczynik, A. (2022). Determination of Anti-Alzheimer's Disease Activity of Selected Plant Ingredients. *Em Molecules* (Vol. 27, Número 10). MDPI. <https://doi.org/10.3390/molecules27103222>

Ul Bari, W., Ur Rehman, N., Khan, A., Halim, S. A., Yuan, Y., Blaskovich, M. A. T., Ziora, Z. M., Zahoor, M., Naz, S., Ullah, R., Alotaibi, A., & Al-Harrasi, A. (2021). Bio-potency and molecular docking studies of isolated compounds from *Grewia optiva* J.R. drumm. ex burret. *Molecules*, 26(7). <https://doi.org/10.3390/molecules26072019>

Venkatesan, K. (2022). Anti-amnesic and anti-cholinesterase activities of  $\alpha$ -asarone against scopolamine-induced memory impairments in rats. *European Review for Medical and Pharmacological Sciences*, 26(17), 6344–6350. [https://doi.org/10.26355/eurrev\\_202209\\_29660](https://doi.org/10.26355/eurrev_202209_29660)

Wagle, A., Seong, S. H., Shrestha, S., Ah Jung, H., & Choi, J. S. (2019). Korean thistle (*cirsium japonicum* var. *Maackii* (Maxim.) Matsum.): A potential dietary supplement against diabetes and Alzheimer's disease. *Molecules*, 24(3). <https://doi.org/10.3390/molecules24030649>

Wan Othman, W. N. N., Liew, S. Y., Khaw, K. Y., Murugaiyah, V., Litaudon, M., & Awang, K. (2016). Cholinesterase inhibitory activity of isoquinoline alkaloids from three *Cryptocarya* species (Lauraceae). *Bioorganic and Medicinal Chemistry*, 24(18), 4464–4469. <https://doi.org/10.1016/j.bmc.2016.07.043>

Wan, Y.-Y., Wang, C.-F., Wang, Q.-H., Xiao, Y., Wang, Z.-B., & Kuang, H.-X. (2016). Study on active constituents against Alzheimer's disease from *Valeriana amurensis*. *Zhongguo Zhongyao Zazhi*, 41(9), 1649–1653. <https://doi.org/10.4268/cjcm20160914>

Wang, A.-W., Liu, Y.-M., Zhu, M.-M., & Ma, R.-X. (2022). Isosteroidal alkaloids of *Fritillaria taipaiensis* and their implication to Alzheimer's disease: Isolation, structural elucidation and biological activity. *Phytochemistry*, 201. <https://doi.org/10.1016/j.phytochem.2022.113279>

Wang, C.-H., Zheng, X.-Y., Zhang, Z.-J., Chou, G.-X., Wu, T., Cheng, X.-M., & Wang, Z.-T. (2009). Acetylcholinesterase inhibitive activity-guided isolation of two new alkaloids from seeds of *Peganum*



nigellastrum Bunge by an in vitro TLC- bioautographic assay. *Archives of Pharmacal Research*, 32(9), 1245–1251. <https://doi.org/10.1007/s12272-009-1910-x>

Wang, Q., Wang, C., Zuo, Y., Wang, Z., Yang, B., & Kuang, H. (2012). Compounds from the roots and rhizomes of valeriana amurensis protect against neurotoxicity in PC12 cells. *Molecules*, 17(12), 15013–15021. <https://doi.org/10.3390/molecules171215013>

Wang, S.-N., Li, Q., Jing, M.-H., Alba, E., Yang, X.-H., Sabaté, R., Han, Y.-F., Pi, R.-B., Lan, W.-J., Yang, X.-B., & Chen, J.-K. (2016). Natural Xanthones from Garcinia mangostana with Multifunctional Activities for the Therapy of Alzheimer's Disease. *Neurochemical Research*, 41(7), 1806–1817. <https://doi.org/10.1007/s11064-016-1896-y>

Wang, X. J., Wang, L. Y., Fu, Y., Wu, J., Tang, X. C., Zhao, W. M., & Zhang, H. Y. (2013). Promising effects on ameliorating mitochondrial function and enhancing Akt signaling in SH-SY5Y cells by (M)-bicyclaphanol A, a novel dimeric podocarpane type trinorditerpene isolated from Celastrus orbiculatus. *Phytomedicine*, 20(12), 1064–1070. <https://doi.org/10.1016/j.phymed.2013.04.017>

Wang, Y.-H., Zhang, Z.-K., Yang, F.-M., Sun, Q.-Y., He, H.-P., Di, Y.-T., Mu, S.-Z., Lu, Y., Chang, Y., Zheng, Q.-T., Ding, M., Dong, J.-H., & Hao, X.-J. (2007). Benzylphenethylamine alkaloids from Hosta plantaginea with inhibitory activity against tobacco mosaic virus and acetylcholinesterase. *Journal of Natural Products*, 70(9), 1458–1461. <https://doi.org/10.1021/np0702077>

Wang, Y.-M., Ming, W.-Z., Liang, H., Wang, Y.-J., Zhang, Y.-H., & Meng, D.-L. (2020). Isoquinolines from national herb Corydalis tomentella and neuroprotective effect against lipopolysaccharide-induced BV2 microglia cells. *Bioorganic Chemistry*, 95. <https://doi.org/10.1016/j.bioorg.2019.103489>

Wang, Y.-X., Ren, Q., Yan, Z.-Y., Wang, W., Zhao, L., Bai, M., Wang, X.-B., Huang, X.-X., & Song, S.-J. (2017). Flavonoids and their derivatives with β-amyloid aggregation inhibitory activity from the leaves and twigs of Pithecellobium clypearia Benth. *Bioorganic and Medicinal Chemistry Letters*, 27(21), 4823–4827. <https://doi.org/10.1016/j.bmcl.2017.09.051>

Wu, Q., Naeem, A., Zou, J., Yu, C., Wang, Y., Chen, J., & Ping, Y. (2022). Isolation of Phenolic Compounds from Raspberry Based on Molecular Imprinting Techniques and Investigation of Their Anti-Alzheimer's Disease Properties. *Molecules*, 27(20). <https://doi.org/10.3390/molecules27206893>  
Xia, W., Luo, P., Hua, P., Ding, P., Li, C., Xu, J., Zhou, H., & Gu, Q. (2019). Discovery of a New Pterocarpan-Type Antineuroinflammatory Compound from Sophora tonkinensis through Suppression of the TLR4/NFκB/MAPK Signaling Pathway with PU.1 as a Potential Target. *ACS Chemical Neuroscience*, 10(1), 295–303. <https://doi.org/10.1021/acscchemneuro.8b00243>

Xian, Y.-F., Lin, Z.-X., Mao, Q.-Q., Hu, Z., Zhao, M., Che, C.-T., & Ip, S.-P. (2012). Bioassay-guided isolation of neuroprotective compounds from uncaria rhynchophylla against beta-amyloid-induced neurotoxicity. *Evidence-Based Complementary and Alternative Medicine*, 2012. <https://doi.org/10.1155/2012/802625>

Xin, L., Yamujala, R., Wang, Y., Wang, H., Wu, W.-H., Lawton, M. A., Long, C., & Di, R. (2013). Acetylcholineesterase-Inhibiting Alkaloids from Lycoris radiata Delay Paralysis of Amyloid Beta-Expressing Transgenic C. elegans CL4176. *PLoS ONE*, 8(5). <https://doi.org/10.1371/journal.pone.0063874>

Xu, S. L., Choi, R. C. Y., Zhu, K. Y., Leung, K.-W., Guo, A. J. Y., Bi, D., Xu, H., Lau, D. T. W., Dong, T. T. X., & Tsim, K. W. K. (2012). Isorhamnetin, a flavonol aglycone from Ginkgo biloba L., induces neuronal differentiation of cultured PC12 cells: Potentiating the effect of nerve growth factor.



*Evidence-Based Complementary and Alternative Medicine*, 2012.  
<https://doi.org/10.1155/2012/278273>

Yan, Y., Ran, X., Wang, D., Zhang, X., Peng, M., Yan, X., Tang, L., Liang, H., Qin, X., Di, Y.-T., Luo, R., Hao, X.-J., & Yao, Y.-G. (2022). Munronin V with 7/7/6 tricarbocyclic framework from Munronia henryi harms inhibits tau pathology by activating autophagy. *Organic and Biomolecular Chemistry*, 21(3), 514–519. <https://doi.org/10.1039/d2ob01965e>

Yang, E.-J., Lee, T., & Song, K.-S. (2019).  $\beta$ -Secretase inhibition by C-methylisoflavones from Abronia nana. *Natural Product Research*, 33(12), 1705–1712. <https://doi.org/10.1080/14786419.2018.1431637>

Yang, X., Peng, Q., Liu, Q., Hu, J., Tang, Z., Cui, L., Lin, Z., Xu, B., Lu, K., Yang, F., Sheng, Z., Yuan, Q., Liu, S., Zhang, J., & Zhou, X. (2017). Antioxidant activity against h2 o2-induced cytotoxicity of the ethanol extract and compounds from pyrola decorate leaves. *Pharmaceutical Biology*, 55(1), 1843–1848. <https://doi.org/10.1080/13880209.2017.1333126>

Yang, Z., Zhang, D., Ren, J., Yang, M., & Li, S. (2012). Acetylcholinesterase inhibitory activity of the total alkaloid from traditional Chinese herbal medicine for treating Alzheimer's disease. *Medicinal Chemistry Research*, 21(6), 734–738. <https://doi.org/10.1007/s00044-011-9582-8>

Yano, M., Nakashima, S., Kasa, S., Nakamura, S., Nishimura, K., Oda, Y., Takata, K., & Matsuda, H. (2020). Accelerative effects of carbazole-type alkaloids from Murraea koenigii on neurite outgrowth and their derivative's in vivo study for spatial memory. *Journal of Natural Medicines*, 74(2), 448–455. <https://doi.org/10.1007/s11418-020-01388-8>

Yano, M., Nakashima, S., Oda, Y., Nakamura, S., & Matsuda, H. (2020). BBB-permeable aporphine-type alkaloids in Nelumbo nucifera flowers with accelerative effects on neurite outgrowth in PC-12 cells. *Journal of Natural Medicines*, 74(1), 212–218. <https://doi.org/10.1007/s11418-019-01368-7>  
Yen, P.-L., Cheng, S.-S., Wei, C.-C., Lin, H.-Y., Liao, V. H.-C., & Chang, S.-T. (2016). Antioxidant activities and reduced amyloid- $\beta$  toxicity of 7-hydroxycalamenene isolated from the essential oil of zelkova serrata heartwood. *Natural Product Communications*, 11(9), 1357–1362. <https://doi.org/10.1177/1934578x1601100943>

Yin, F., Sancheti, H., Patil, I., & Cadena, E. (2016). Energy metabolism and inflammation in brain aging and Alzheimer's disease. In *Free Radical Biology and Medicine* (Vol. 100, p. 108–122). Elsevier Inc. <https://doi.org/10.1016/j.freeradbiomed.2016.04.200>

Yoshioka, T., Murakami, K., Ido, K., Hanaki, M., Yamaguchi, K., Midorikawa, S., Taniwaki, S., Gunji, H., & Irie, K. (2016). Semisynthesis and structure-activity studies of uncarinic acid C isolated from uncaria rhynchophylla as a specific inhibitor of the nucleation phase in amyloid  $\beta$ 42 aggregation. *Journal of Natural Products*, 79(10), 2521–2529. <https://doi.org/10.1021/acs.jnatprod.6b00392>  
Yousof Ali, M., Jung, H. A., & Choi, J. S. (2015). Anti-diabetic and anti-Alzheimer's disease activities of Angelica decursiva. *Archives of Pharmacal Research*, 38(12), 2216–2227. <https://doi.org/10.1007/s12272-015-0629-0>

Yu, Y., Zhou, L., Sun, M., Zhou, T., Zhong, K., Wang, H., Liu, Y., Liu, X., Xiao, R., Ge, J., Tu, P., Fan, D. S., Lan, Y., Hui, C., & Chui, D. (2012). Xylocoside G reduces amyloid- $\beta$  induced neurotoxicity by inhibiting NF- $\kappa$ B signaling pathway in neuronal cells. *Journal of Alzheimer's Disease*, 30(2), 263–275. <https://doi.org/10.3233/JAD-2012-110779>

Zhan, G., Miao, R., Zhang, F., Chang, G., Zhang, L., Zhang, X., Zhang, H., & Guo, Z. (2020). Monoterpene indole alkaloids with acetylcholinesterase inhibitory activity from the leaves of Rauvolfia vomitoria. *Bioorganic Chemistry*, 102. <https://doi.org/10.1016/j.bioorg.2020.104136>



Zhang, L., Yu, H., Zhao, X., Lin, X., Tan, C., Cao, G., & Wang, Z. (2010). Neuroprotective effects of salidroside against beta-amyloid-induced oxidative stress in SH-SY5Y human neuroblastoma cells. *Neurochemistry International*, 57(5), 547–555. <https://doi.org/10.1016/j.neuint.2010.06.021>

Zhang, X. D., Liu, X. Q., Kim, Y. H., & Whang, W. K. (2014). Chemical constituents and their acetyl cholinesterase inhibitory and antioxidant activities from leaves of Acanthopanax henryi: Potential complementary source against Alzheimer's disease. *Archives of Pharmacal Research*, 37(5), 606–616. <https://doi.org/10.1007/s12272-013-0252-x>

Zhang, X., Wang, Y., Qin, Q., Wang, Y., Xu, J., & He, X. (2021). Pronounced anti-neuroinflammatory jasmonates and terpenes isolated from lychee seeds. *Fitoterapia*, 152. <https://doi.org/10.1016/j.fitote.2021.104924>

Zhao, D., Gu, M.-Y., Zhang, L. J., Jeon, H. J., Cho, Y.-B., & Yang, H. O. (2019). 7-Deoxy- trans-dihydronarciclasine Isolated from Lycoris chejuensis Inhibits Neuroinflammation in Experimental Models. *Journal of Agricultural and Food Chemistry*, 67(35), 9796–9804. <https://doi.org/10.1021/acs.jafc.9b03307>

Zhao, G., Yao-Yue, C., Qin, G.-W., & Guo, L.-H. (2012). Luteolin from Purple Perilla mitigates ROS insult particularly in primary neurons. *Neurobiology of Aging*, 33(1), 176–186. <https://doi.org/10.1016/j.neurobiolaging.2010.02.013>

Zhao, X., Liao, Z., Qi, Y., Shen, X., Bi, K., & Jia, Y. (2016). Antioxidative activity of methyl amygdalinate from the seeds of: Prunus persica and neuroprotective effects on A $\beta$ 1-42-induced neurodegeneration models. *RSC Advances*, 6(96), 93794–93800. <https://doi.org/10.1039/c6ra18913j>

Zhou, Z.-Q., Fan, H.-X., He, R.-R., Xiao, J., Tsoi, B., Lan, K.-H., Kurihara, H., So, K.-F., Yao, X.-S., & Gao, H. (2016). Lycobarbarspermidines A-O, New Dicaffeoylspermidine Derivatives from Wolfberry, with Activities against Alzheimer's Disease and Oxidation. *Journal of Agricultural and Food Chemistry*, 64(11), 2223–2237. <https://doi.org/10.1021/acs.jafc.5b05274>

Zhumanova, K., Lee, G., Baiseitova, A., Shah, A. B., Kim, J. H., Kim, J. Y., Lee, K. W., & Park, K. H. (2021). Inhibitory mechanism of O-methylated quercetins, highly potent  $\beta$ -secretase inhibitors isolated from Caragana balchaschensis (Kom.) Pojark. *Journal of Ethnopharmacology*, 272. <https://doi.org/10.1016/j.jep.2021.113935>

Zou, M., Wang, R., Yin, Q., & Liu, L. (2021). Bioassay-guided isolation and identification of anti-Alzheimer's active compounds from Spiranthes sinensis (Pers.) Ames. *Medicinal Chemistry Research*, 30(10), 1849–1855. <https://doi.org/10.1007/s00044-021-02777-8>